Imaging Across Biological Length Scales with the Emerging Frontiers in Microscopy at HHMI Janelia Research Campus

Advanced Imaging Center Howard Hughes Medical Institute Janelia Research Campus

Resolution limit

Many thousands of proteins can fit within a single diffraction limited spot!

300nm

One GFP Molecule



2-4nm

Proposed method for molecular optical imaging

E. Betzig

NSOM Enterprises, 17 Webster Drive, Berkeley Heights, New Jersey 07922

Received September 20, 1994

We can resolve multiple discrete features within a focal region of m spatial dimensions by first isolating each on the basis of $n \ge 1$ unique optical characteristics and then measuring their relative spatial coordinates. The minimum acceptable separation between features depends on the point-spread function in the (m + n)-dimensional space formed by the spatial coordinates and the optical parameters, whereas the absolute spatial resolution is determined by the accuracy to which the coordinates can be measured. Estimates of each suggest that near-field fluorescence excitation microscopy/spectroscopy with molecular sensitivity and spatial resolution is possible.





Interferometric PALM system at the AIC

Newport

Dell

. . . .

Interferometric PALM (iPALM)





Z = 0 nm

Courtesy of Dr. Johanna Ivaska University of Turku, Finland iPALM at AIC





Resolving the inner and outer membranes of a single bacterium on iPALM

Courtesy of Dr. Phil Klebba Kansas State University iPALM at AIC



Pros and Cons of Current Super-Resolution Techniques

Nyquist Criterion:

N -fold resolution increase in **D** dimensions $\rightarrow N^{P}$ -fold more photons collected





Light sheet/plane illumination microscopy



Widefield light collection lens No photon wasted due to pinhole rejection



Only the plane that is being imaged is illuminated



Temporal resolution: 800 x 400 x 200 µm³ at **5 Hz** Spatial resolution: **400 nm** laterally, **1.8 µm** axially 175 million voxels per second



Neuronal signaling in whole zebrafish brain

Misha Ahrens and Philipp Keller HHMI Janelia Research Campus Nature Methods 10: 413-420, 2013

Imaging high scattering sample: Drosophila embryogenesis



Philipp Keller HHMI Janelia Research Campus



Optical challenges in light-sheet imaging of large specimens Spat

Spatial mismatches in illumination and detection processes

Simultaneous MultiView (SiMView) light sheet microscope at the AIC



Intensity 100 µm

00:04:00

Adaptive imaging of mouse post-implantation development

Projection of imaging volume

Real-time optimization of imaging conditions

Visualizing early heart formation





00:00:00

Nkx2.5-Cre; R26^{mT/mG}



myr-eGFP



Putting it all together: vector flow of every cell in mouse embryo





00:40:00

Lateral view





Gaussian beam profile during propagation

Bessel beam profile during propagation

Gaussian beam profile during propagation





Bessel beam profile during propagation



Lattice light sheet microscope at the AIC





400 stacks, 1.27s/2-color stack



Courtesy of Dr. Lakshmi Bagolapalan NIH Lattice light sheet microscope at AIC

En face View

Mitochondrial segregation during HeLa cell division

Courtesy of Dr. Erika Holzbaur University of Pennsylvania Lattice light sheet microscope at Janelia



Synchronized cell division in Drosophila

Courtesy of Dr. Edouard Bertrand Lattice light sheet microscopy at the AIC IGMM-CNRS, Montpelier, France



The hurdle of imaging with increasing penetration depth





Light scattering

Adaptive Optics





170 X 185 X 135 µm 96 hpf zebrafish embryo



MOSAIC: Multimodal Optical System with Adaptive Imaging Correction



Operational modes:

- Gaussian or lattice light sheet
- TPE Gaussian or Bessel light sheet
- Multifocal photon reassignment
- 3D structured illumination
- 3D single molecule localization
- TPE point or Bessel resonant scan
- TPE photon reassignment
- 3D phase imaging
- Expansion lattice light sheet
- Patterned photostimulation

All with adaptive optics



Adaptive optics-lattice light sheet microscopy Eric Betzig (Janelia) & Tomas Kirchhausen (Harvard)

Zebrafish 14-16 hfp

Courtesy of: Drs Eric Betzig, *Janelia Research Campus,* and Tomas Kirchhausen & Gokul Upadhyayula, *Harvard Medical School*

membrane-Citrine cis-golgi: gm130-mNeonGreen ER: sec61-tRFP-t MitoTracker: Deep Red FM Dye

Focused Ion Beam SEM at the AIC

Remove ~2-10 nm with Ga Ion Beam

Image with Scanning electron microscope

Repeat...

But commercial FIB-SEMs have severe limitations:

Interrupts: Destructive and Unforgiving

FIB Source reheat ~ 3 days Room Temperature Fluctuations ~ weekly Utility failure ~ monthly Component Failure ~ quarterly

Restart: Transient Data Loss

Uneven milling Defocus loss



Focused Ion Beam SEM at the AIC

Custom Control Acquisition Hardware/Software Pausing on any system issue e.g.: Temperature excursion, Milling current jump, Focus shift, etc. Long term optimization: Focus, Stigmation, Drift Automated Seamless Restart: Predictive correction of restart transients Enables nights sleep, no data degradation or loss

Cross integrate best technologies: SEM and FIB columns from different vendors

Custom Feedback on ion beam milling current nm position control on milling Automated seamless restart

Stabilized and Fault Tolerant Room Environment, Utilities



HeLa Cell (manual segmentation) 1.2 μ m x 1.2 μ m x 0.9 μ m volume (1.27 μ m³)

1.2 μm x 1.2 μm x 0.9 μm volume (1.27 μm 300 x 300 x 225 pixels



4 nm x 4 nm x 4 nm voxels



Oocyst of *Plasmodium falciparum* inside the mosquito gut

Courtesy of Dr. Flaminia Catteruccia FIB-SEM at the AIC Harvard University





Visitor Program at AIC

Cost of experiment covered by AIC Lodging cost will be 100% covered by HHMI Janelia

Advanced Imaging Center Total Programmatic Support

How the AIC maintains ~85% experimental success rate

Technical consultation <u>AIC</u>

IACUC/IBC review IBC, Vivarium, EH&S, Cell & Mol. Biol. Facilities

Travel arrangement/visa application Lab Coordinator, Visitor Program

> Sample shipment and receiving EH&S, Cell Culture Facility

> > Lodging Visitor Program<u>, Lab Coordinator</u>

Vivarium/model organism handling Vivarium, Drosophila Resources Pre-arrival sample preparation Cell Culture, Histology Facilities

cDNAs with FPs/model organisms Molecular Biology Facility/Various labs

Imaging probes Luke Lavis and Loren Looger's Labs

Microscope time

Imaging assistance AIC

Image processing and analysis AIC

Call-for-Proposals Announced ~ twice a year

Two-tier Review Process

First Tier:Technical assessment by the AIC team
Pass/Fail grade will be assigned
Can the question be addressed by commercial instruments?
Can the AIC instrument(s) deliver the expected results?
Can the specimen be safely handled at Janelia?

Second Tier: Peer Review by HHMI and external reviewers and Moore Foundation Tiered priority score will be assigned How well the scientific rationale supports the project How the experimental question justifies the use of the AIC instruments Discussion of potential problems and alternative approaches Quantitative approach Amount of time required in the AIC to meet the project goals Personnel





Africa Microscopy Initiative





Response to the first proposal call from the AMI Imaging Centre



63 proposals from 18 countries









AIC global microscopy outreach programs

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Various workshops in the US
Canadian Microscopy Workshop
Imaging Latin America
Imaging Africa
Danish Bioimaging Workshop
Croucher Advanced Imaging Course
Okinawa Workshop for Japan & Southeast Asia
RMS/KCL Imaging Bootcamp
Scandivanian Bioimaging Workshop

Israel Microscopy Workshop

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