

# Translating Scientific Breakthroughs from the Genomics Revolution



**Preethi Gunaratne, Ph.D.**

John & Rebecca Moores Professor  
Department of Biology & Biochemistry,  
Director, University of Houston Sequencing Core  
University of Houston

**SEASR 2024 Annual Meeting  
Nashville, TN**

# The University of Houston Sequencing and Gene Editing Core

"Student Run, Comprehensive, Sample to Data Insight Pipelines for Commercial and Academic User Groups"

## Baylor Human Genome Sequencing Center – 2000



- Human Chromosomes  
3, 12, X
- 15,000 FL-cDNA Sequences  
Mammalian Gene Collection

1999

2020

2024



- ~1000 people
- ~300 Instruments
- 9 Months
- \$1B
- 1 Human Genome

**UHSEQ**  
Sequencing Core



### Illumina NextSeq 2000

- Effort of 1 Person
- 1 Instrument
- 48 hrs.
- \$1000/Genome
- 4 Human Genomes

### Illumina NovaSeq X+

- Effort of 1 Person
- 1 Instrument
- 24 hrs.
- \$200/Genome
- 4 Human Genomes

### Illumina NovaSeq X+





## **“Niche Sequencing”**

### **microRNA**

1. Developing microRNA biomarkers and therapeutics for cancer
2. Discovering microRNA-based blood biomarkers for predicting individuals at high risk for Opioid Overuse Disorder (OUD)

### **Non-Canonical Dark Matter**

Platform for designing peptide & mRNA vaccines from immunogenic neoantigens from RNA fusions for the treatment & prevention of cancer

### **16S Sequencing**

A Field-Tested Platform for Extracting Personalized DNA Biomarkers for Predicting Drainage Heights, % Contribution, Well-to-Well Communication for Precision Well Placement  
Providing Metrics for Asset Development



# Comprehensive, Sample to Data Insight Pipelines for Commercial Academic and User Groups



**Preethi Gunaratne, Ph.D.**  
**Director**



**Nuwan Acharige, Ph.D.**



**Lead Scientist**  
**16S Sequencing Platform**

**Chathurani Ranatunge, Ph.D.**



**Lead Analyst**  
**Breast Cancer Prevention**  
**Vaccine Platform**

**Sakuni Rankothgedera**



**Lead Scientist**  
**Single Cell and Spatial**  
**Transcriptomics Platform**

**Shiyanth Thevasagayampillai**



**Lead Scientist**  
**mRNA Vaccine**  
**Discovery Platform**

**Aaranyah Kandasamy**



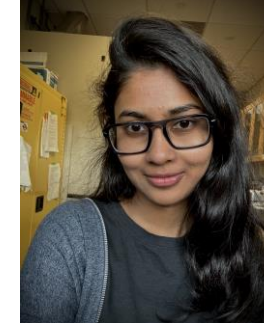
**Lead Scientist**  
**PDX -Actionable Mutation**  
**Discovery Platform**

**Dilshan Adikari**



**NGS Specialist**  
**RNA & DNA**  
**Sequencing Platform**

**Nirmali Samarakoon**



**NGS Specialist**  
**RNA & DNA**  
**Sequencing Platform**

**Enes Dasdemir**



**Single Cell Sequencing**  
**Hematologic Cancers**

**Cole Woody**



***In silico* fusion**  
**validation platform**





**Ashley Benham-Duret, Ph.D.**  
Senior Scientist, Sales Executive II,



**Asha Palat, Ph.D.**  
Scientist,  
Novogene



**Brandon Mistretta, Ph.D.**  
Field Application  
Scientist,



**Ian Wilson, M.Sc.**  
Scientist,



**Micah Castillo, Ph.D.**  
Spatial Field Application  
Scientist



**Kimberly Holloway,  
Ph.D.**  
Associate Director,  
Translational Oncology,  
Iterion Therapeutics

**Sujash Chatterjee, PhD**  
Lead-Product Marketing &  
Commercialization, Genomics,



**Sujash Chatterjee, PhD**  
BioPharma Strategy and  
Commercialization



**Jignesh Chandrana, M.Sc.**  
Lead Lab Automation Developer  
Invitae Genetic Testing

**“We provide custom solutions to your problems and perform optimization until we crack the code”**

- **Industrial Sequencing**
- **Commercial Sequencing**
- **Health Care**
- **Academic Labs**



THE UNIVERSITY OF TEXAS  
**MD Anderson**  
~~Cancer Center~~

 **UTHealth**  
Houston

**Baylor**  
College of  
Medicine

 **THE TEXAS HEART INSTITUTE®**

 **Health**

HOUSTON  
**Methodist**  
LEADING MEDICINE

UNIVERSITY of  
**HOUSTON**

 **TEXAS A&M UNIVERSITY**  
**HEALTH**

 **Texas Children's**  
**Hospital®**

 **TEXAS TECH**  
UNIVERSITY.

 **AUGUSTA**  
UNIVERSITY

 **LEHIGH**  
UNIVERSITY

 **Queen's**  
UNIVERSITY

**UF**  
UNIVERSITY of  
**FLORIDA**

 **Cleveland Clinic**

 **NEW YORK UNIVERSITY**

 **COLUMBIA COLLEGE**  
COLUMBIA UNIVERSITY in the CITY of NEW YORK



# Industrial Clients



**PETRONAS**

xen<sup>o</sup>START

**GeOME**Analytics





# The University of Houston Sequencing and Gene Editing Core **UHSEQ**

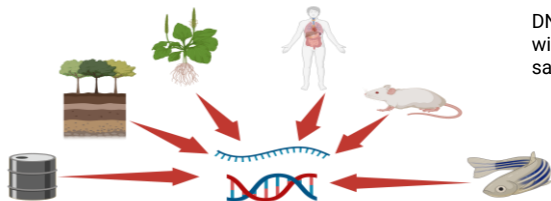
Sequencing Core

"Student Run, Comprehensive, Sample to Data Insight Pipelines for Commercial and Academic User Groups"

## Experimental Design and Assay Optimization

- Consult with our team prior to each assay to ensure optimal results.
- Streamline your projects to avoid future repeats

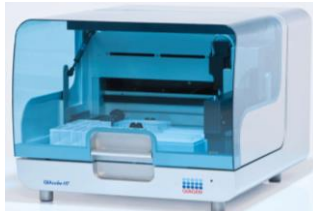
## Sample Preparation and Nuclei Acid Extraction



DNA/RNA Extraction from a wide variety of fresh/preserved sample types.



**QIAcube**: 12 samples/hour  
Compatibility: DNA/RNA



**QIAcube HT**: 96 samples/hour  
Compatibility: DNA/RNA

## Sample Quality Assessment



### Qubit Flex Fluorometer

Throughput: 8 samples/run, 96 samples/10 minutes  
Compatibility: DNA/RNA Protein  
Application: Sample Concentration

### Agilent 4200 TapeStation System

Throughput: 96 samples/1.5 hours  
Compatibility: DNA/RNA  
Application: Sample Quality, Size distribution, RINe, DV200

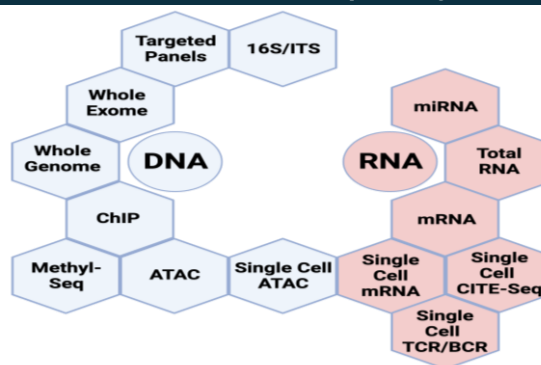
## Multi-Assay RT-PCR

### Agilent Aria MX Real Time PCR System

Throughput: 96 samples  
Compatibility: SYBR/ROX/FAM  
Application: Sample Concentration, Gene Expression Detection, Mouse/Human Tissue Contamination Detection (PDX)



## Next Generation Sequencing



### Illumina MiSeq

Maximum Read Output: 25M  
Maximum Cycle Output: 600



### Illumina NextSeq 2000

Maximum Read Output: 1.1B  
Maximum Cycle Output: 600

## Single Cell Sequencing



### 10X Genomics Instrumentation

A) **Chromium Controller**: 3'/5' Gene Expression, Immune Profiling, ATAC, CITE-Seq

B) **Chromium X**: Fixed RNA, High-throughput, All other assays.

C) **Visium CytAssist**: Spatial profiling of RNA and Protein on FFPE or Fresh-Frozen tissue slides.

D) **Chromium Connect**: Full automation of 10X pipeline.

## Industrial Collaborations

Cancer

xen<sup>o</sup>START

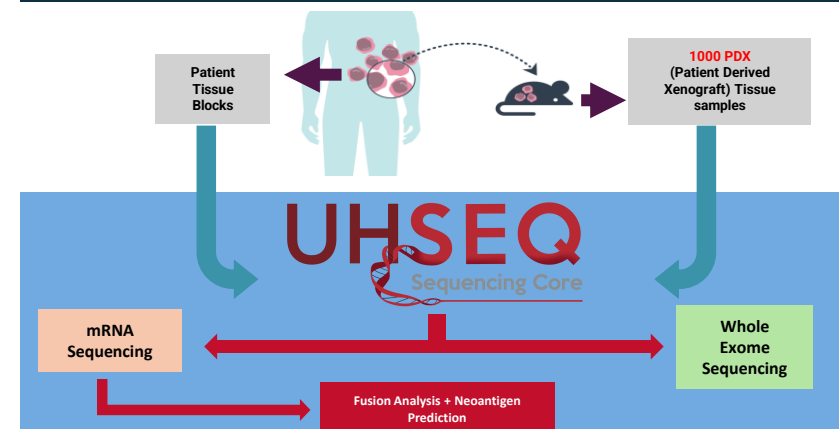
GeOME ANALYTICS

Energy

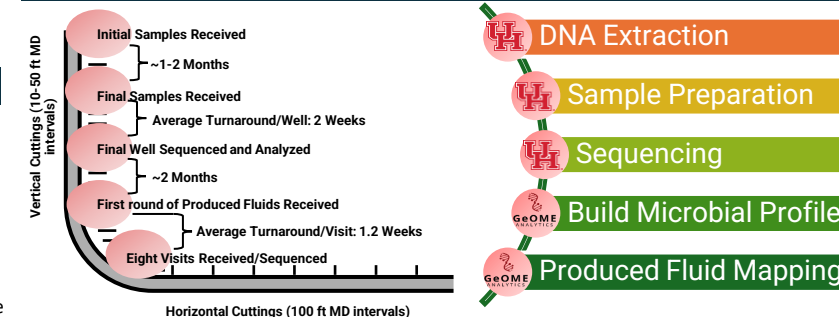
Chevron



## Actionable Mutations in PDX Models



## Reservoir Drainage Diagnostics



## Auxiliary Instrumentation



**Countess 3 FL**  
Compatibility: RFP + GFP  
Application: Cell Counting

**Transblot Turbo Transfer System**  
Application: Western Blotting



## Funding

- Large Core Equipment Grant – 2021
- Large Core Equipment Grant Program -2022

## Genomics

- Whole Genome Seq
- Whole Exome Seq
- 16S rRNA Seq
- ITS Seq
- Metagenomics



## Single-cell Sequencing

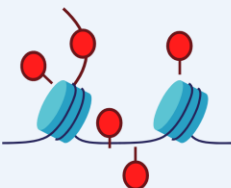
- Single-Cell Gene Expression (3',5' GEX)
- Single-Cell Immune Profiling (5' VDJ)
- Single-Cell ATAC
- Single-Cell Multiome ATAC + GEX
- Single-Cell Gene Expression Flex

## Bioinformatics Analysis

Analysis available for all of the services provided. Our team can create custom analyses tailored to your project goals upon request

## Epigenomics

- ChIP-seq
- ATAC-seq
- Hi-C Assay
- Methyl Seq



# SERVICES WE PROVIDE

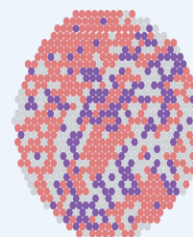
## Transcriptomics

- mRNA-seq
- Total RNA-seq
- miRNA-seq



## Spatial Transcriptomics

- Visium CytAssist
- Visium HD

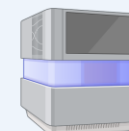


## Sequencing Only

Sequence pre-made libraries with us!



miSeq

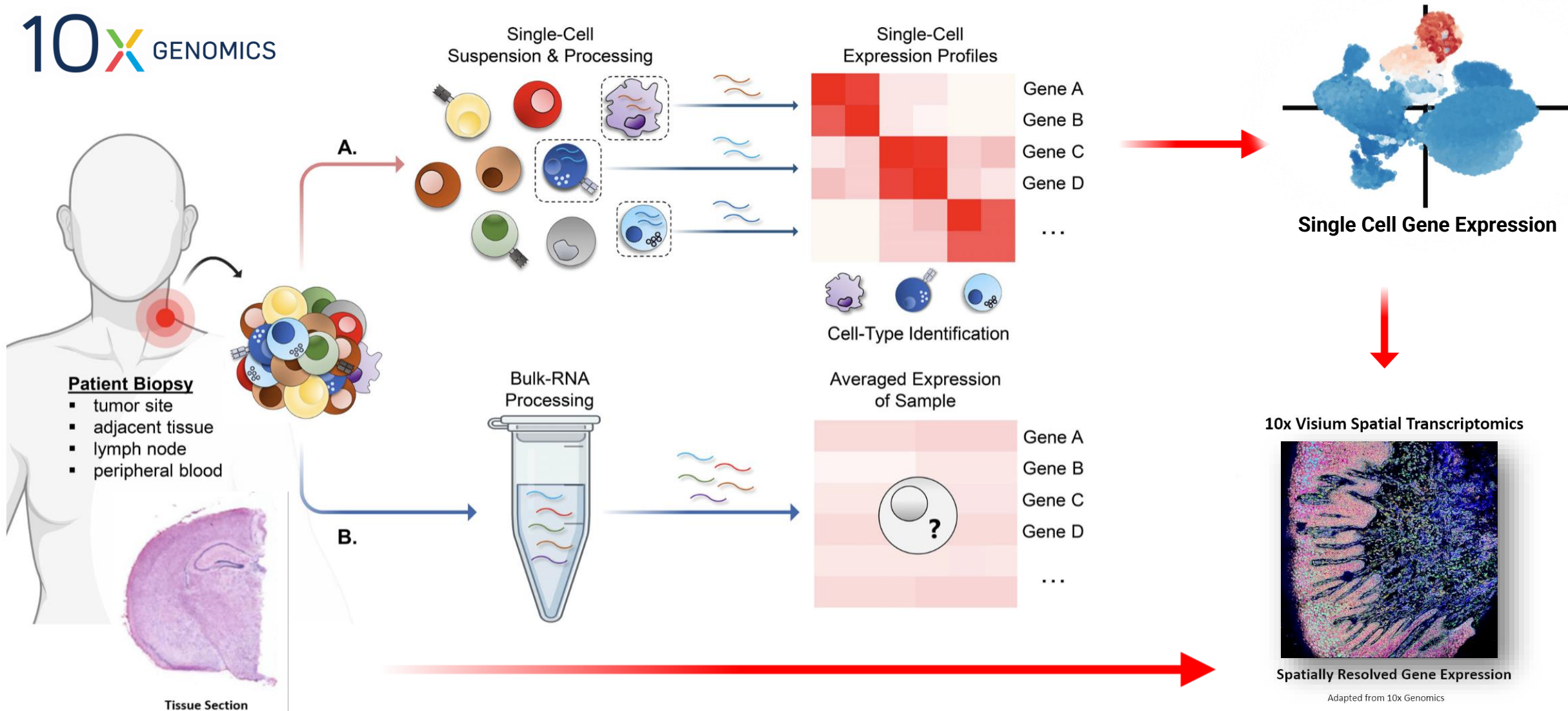


Nextseq 2000



# Bulk RNA Seq Vs Single Cell RNA Seq

10x GENOMICS



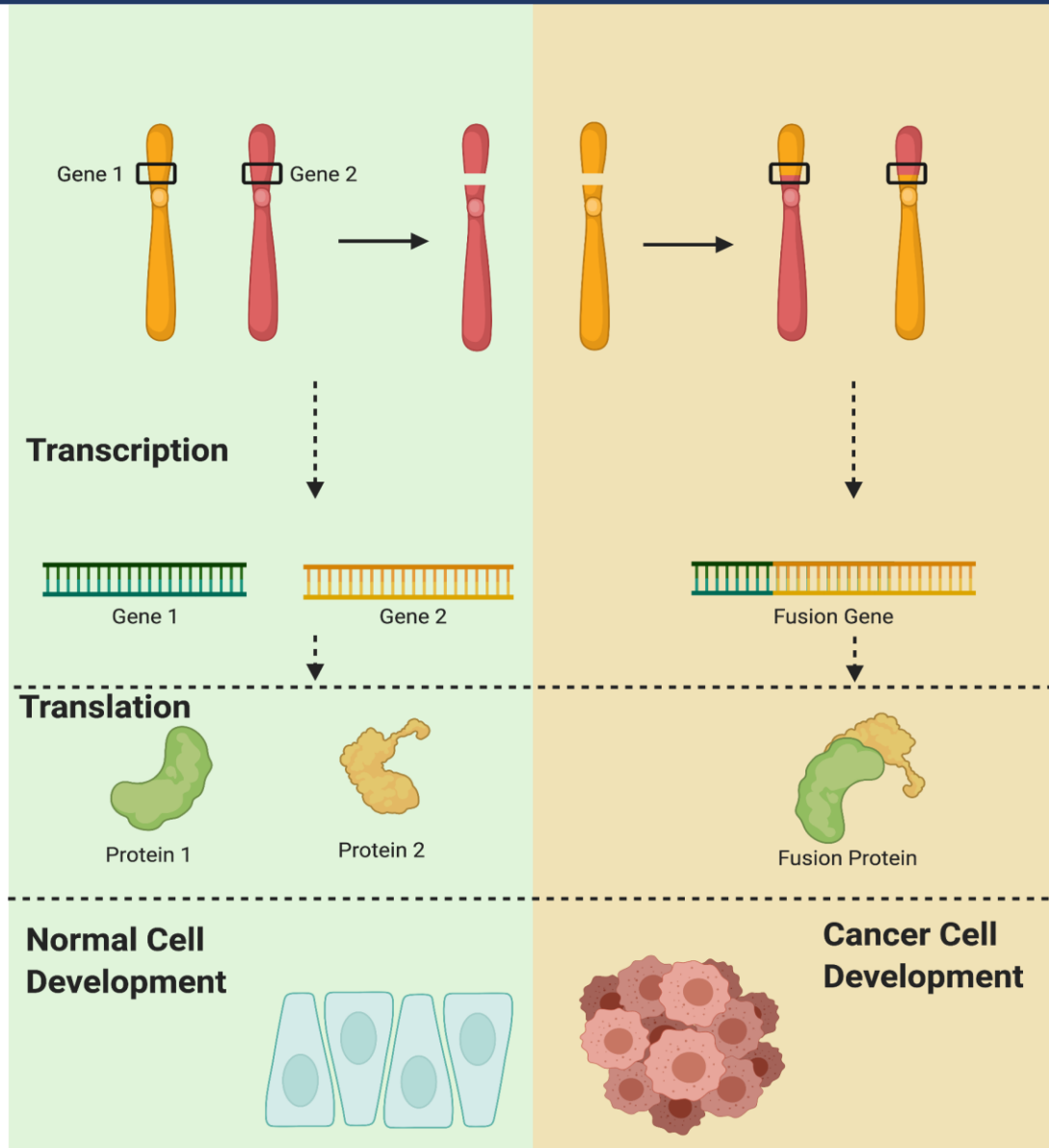
# Discovery of Immunogenic Neopeptides from Actionable Fusions

## to Develop Peptide & mRNA Vaccines for Cancer Treatment &



Micah Castillo, Ph.D.

# Gene Fusions



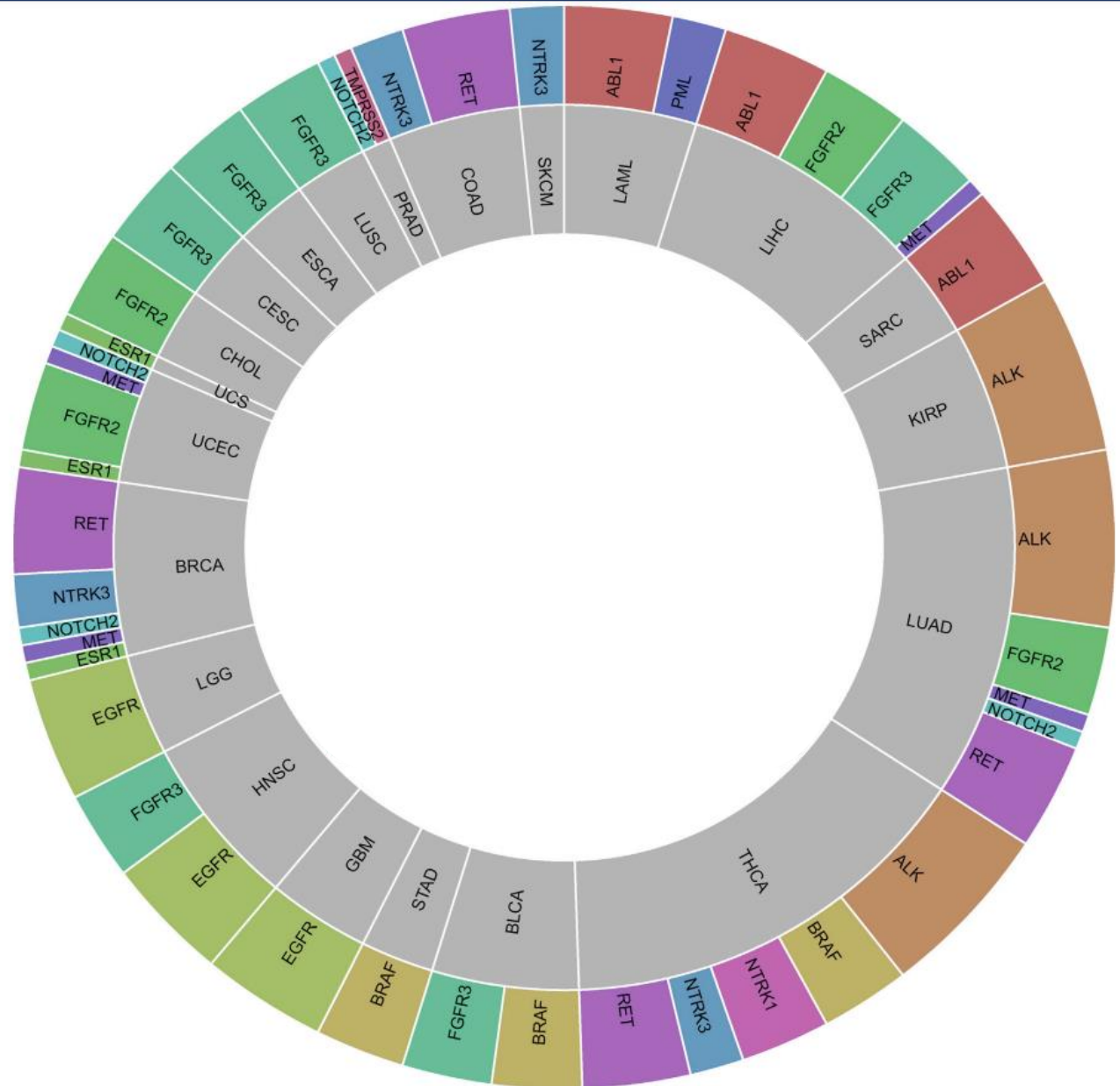
## Importance in Cancer

- Structural chromosome rearrangements result in the exchange of coding or regulatory DNA sequences between genes.
- Fusions may also occur on the RNA level as a result of aberrant splicing.
- Many fusions are strong driver mutations in neoplasia.
- **BCR-ABL fusion gene is a major marker in chronic myelogenous leukemia (CML).**
- Identifying fusions may lead to development of novel therapeutics or early detection methods.

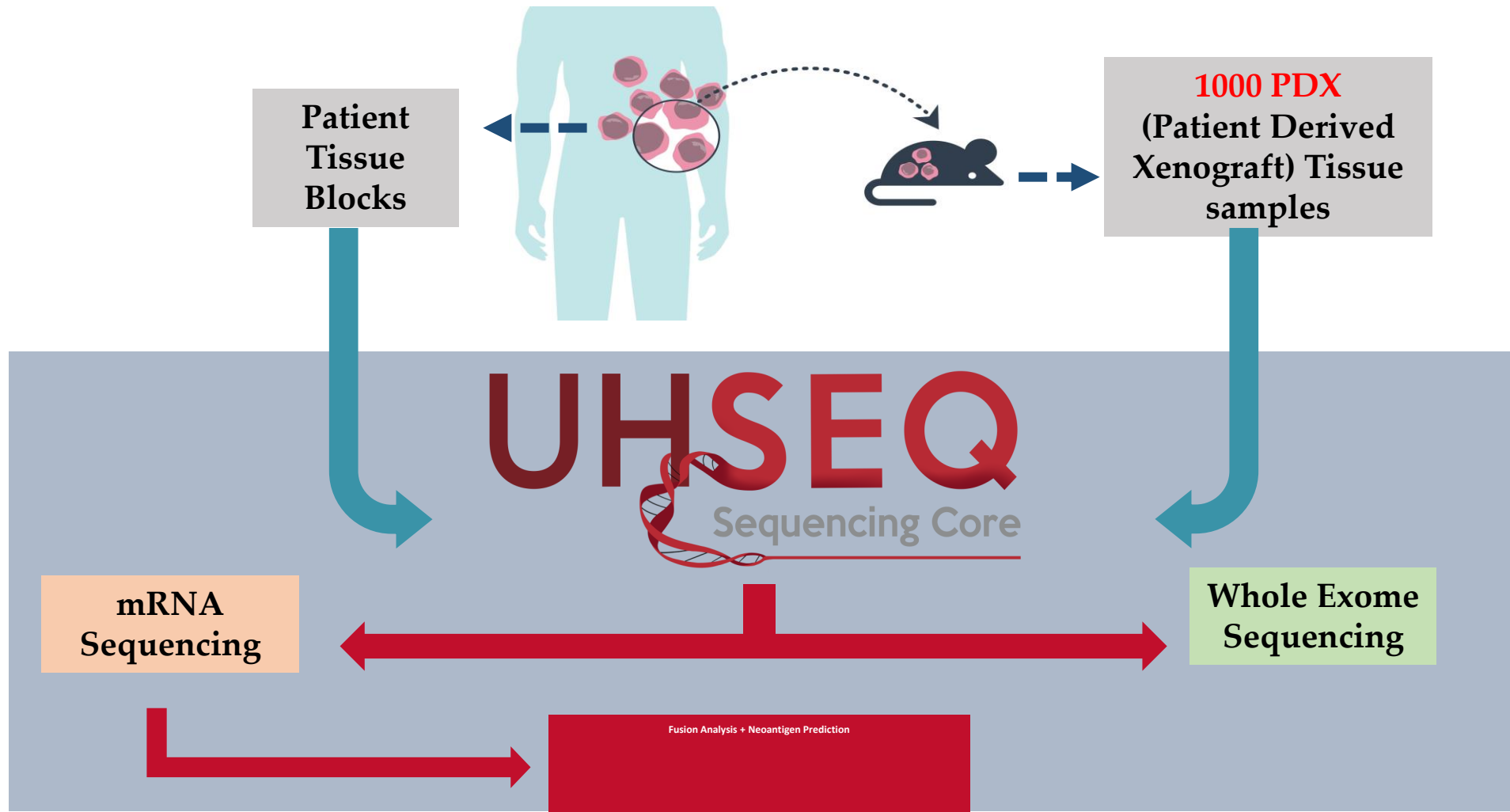


# TCGA Actionable Fusions List

Gene	Drug(s)	Fusion Partner(s)
ABL1	Bosutinib, Dasatinib, Imatinib, Nilotinib, Ponatinib, Venetoclax	FUBP3, BCR
ALK	Alectinib, Brigatinib, Ceritinib, Crizotinib, Entrectinib, Lorlatinib, MEK inhibitors, novel ALK inhibitors, PF2341066, TAE684	EML4, STRN
BRAF	Cobimetinib, MEK inhibitors, Selumetinib, Sorafenib, Trametinib	MRPS33, SND1
EGFR	Afatinib, EGFR TKIs, Erlotinib, first-generation and irreversible EGFR TKIs, gefitinib, HSP90 inhibitors, ZD6474	SEPT14
ESR1	Anti-estrogens	CCDC170, TMEM2
FGFR2	AZD4547, BGJ398, Debio1347, Erdafitinib, FGFR inhibitors	ATE1, BICC1, SHTN
FGFR3	AZD4547, BGJ398, Debio1347, Erdafitinib, FGFR inhibitors	TACC3
MET	Crizotinib	CAV1, ST7
NOTCH2	Gamma secretase inhibitors	SEC22B
NTRK1	Crizotinib, Entrectinib, IGF1R inhibitors, Larotrectinib, pan-TRK inhibitor	IRF2BP2, TPM3
NTRK3	Entrectinib, Larotrectinib, Midostaurin	ETV6
PML	Arsenic Trioxide, Tretinoin, Volasertib	RARA
RET	BLU-667, Crizotinib, LOXO-292, Nintedanib, Sunitinib, Vandetanib	CCDC6, ERC1, NCOA
TMPRSS2	DNA-PKc inhibitors	ERG, ETV4



# Actionable Mutations in PDX Models



# Discordant Reads

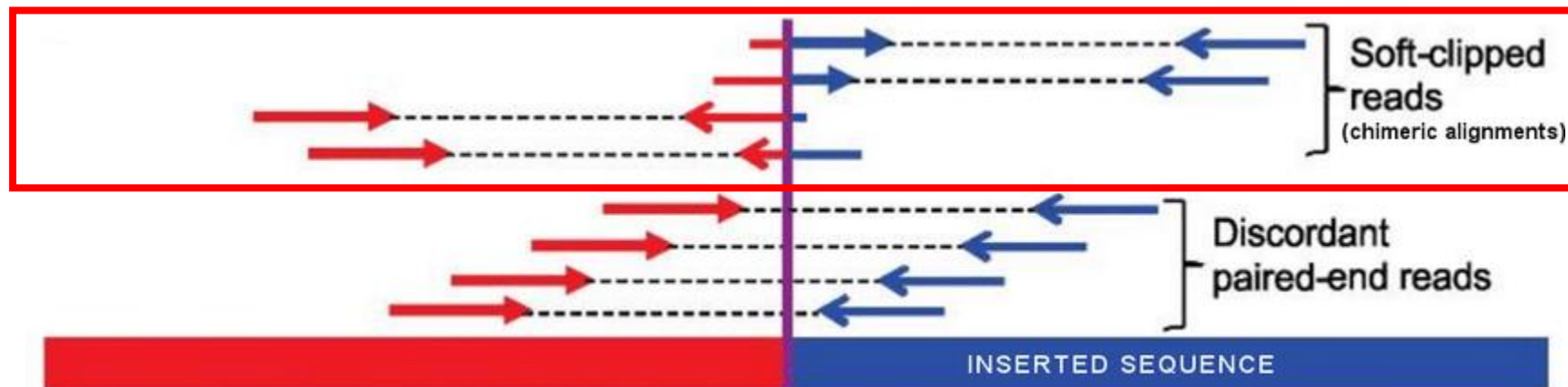
Paired-end reads with unexpected mapping distances.

## Fusion callers that use only discordant reads

- **VariantMeta Caller (DNA fusions)**
  - Gezsi et al. VariantMetaCaller: automated fusion of variant calling pipelines for quantitative, precision-based filtering. *BMC Genomics*. 2015
- **ChimeRScope**
  - Li et al. ChimeRScope: a novel alignment-free algorithm for fusion transcript prediction using paired-end RNA-Seq data. *Nucleic Acids Research*. 2017
- **ChimPipe**
  - Rodriguez-Martin et al. ChimPipe: Accurate detection of fusion genes and transcription-induced chimeras from RNA-seq data. *BMC Genomics*. 2018

## 50% False Discovery Rate

- Mismapping of read pairs to different gene family members and pseudogenes
- Mismapping reads containing repeats shared by two different genes





# Chimeric RNA Discovery Pipeline using Multiple Fusion Callers

**Library Preparation of NSCLC Patient Samples and Next Generation Sequencing**

RNA Seq Data from Lung Cancer Patient Samples



**Identification of gene fusions through CLC Genomics Workbench 20, Dragen Fusion Caller, and EasyFuse Fusion Caller**

Aligned to Human Reference  
Genome Hg38

Detect Fusion Gene tool re-  
mappings the unaligned ends of  
reads (CLC)

Refine Fusion tool re-counts the  
number of fusion crossing  
reads (CLC)



**Filter out Fusions Overlapping in Adjacent Normal Samples**

Subtract fusions represented in both normal and tumor patient samples

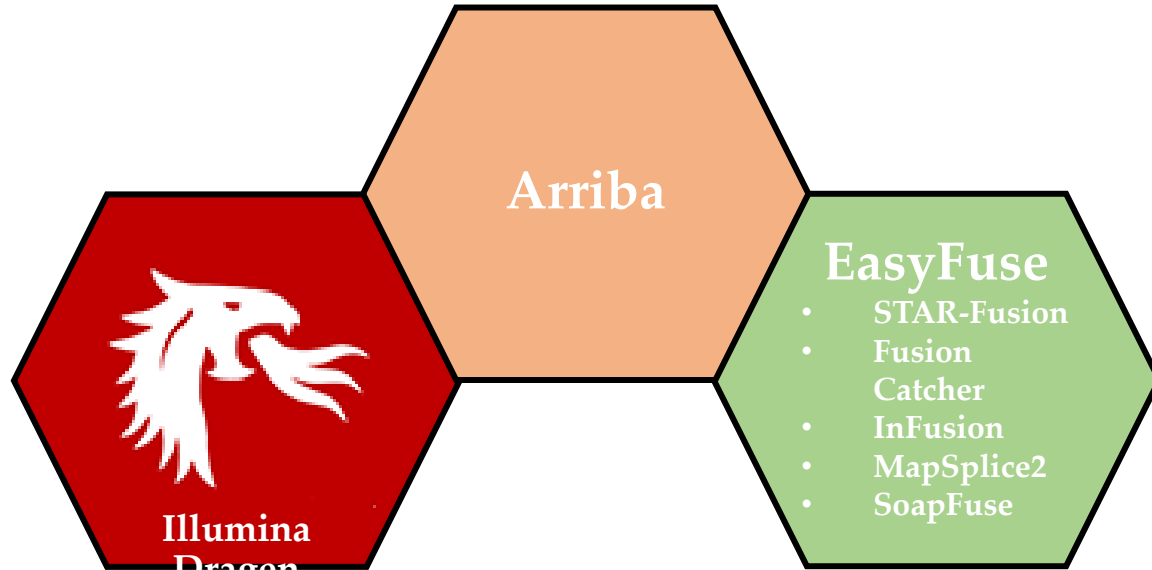


**Overlap with TCGA dataset to assess actionability**

Novel Fusions reported by a 1000 sample TCGA Breast Cancer Dataset run by the University of Chicago's fusion caller.

# Fusion Discovery Pipeline

Higher Stringency, Higher Specificity



Arriba - Uhrig *et al.*, 2021, (*Genome Research*)  
EasyFuse - Weber, *et al.*, 2022, (*Nature Biotechnology*)

Lower Stringency, Higher Sensitivity



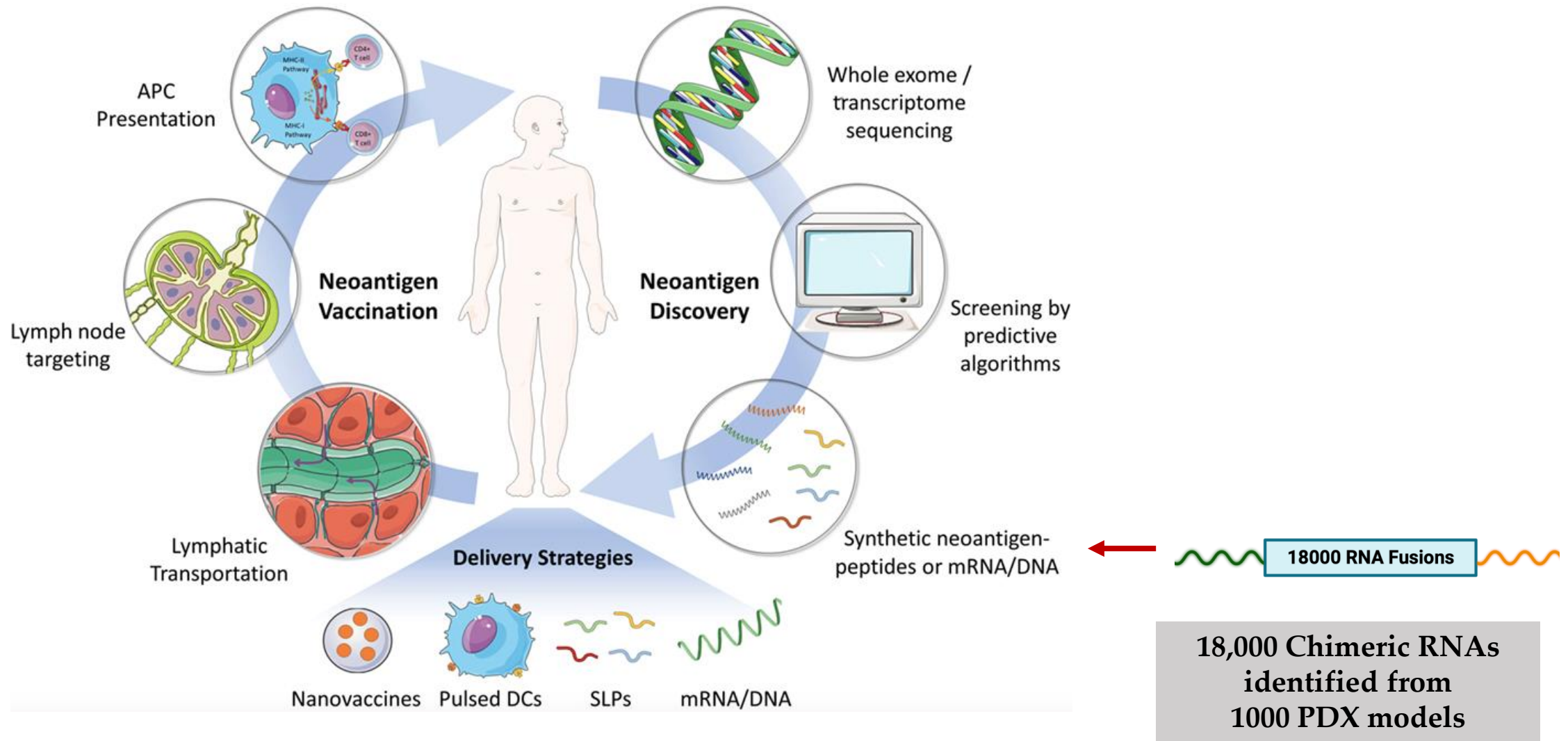
- QIAGEN CLC Genomics Workbench 20 Fusion Caller
  - Detects split-reads
  - Extra alignment step to artificially generated reference
    - Increase in the number of reads confidently mapped across fusion junctions

Overlap with TCGA Actionable Fusion Genes

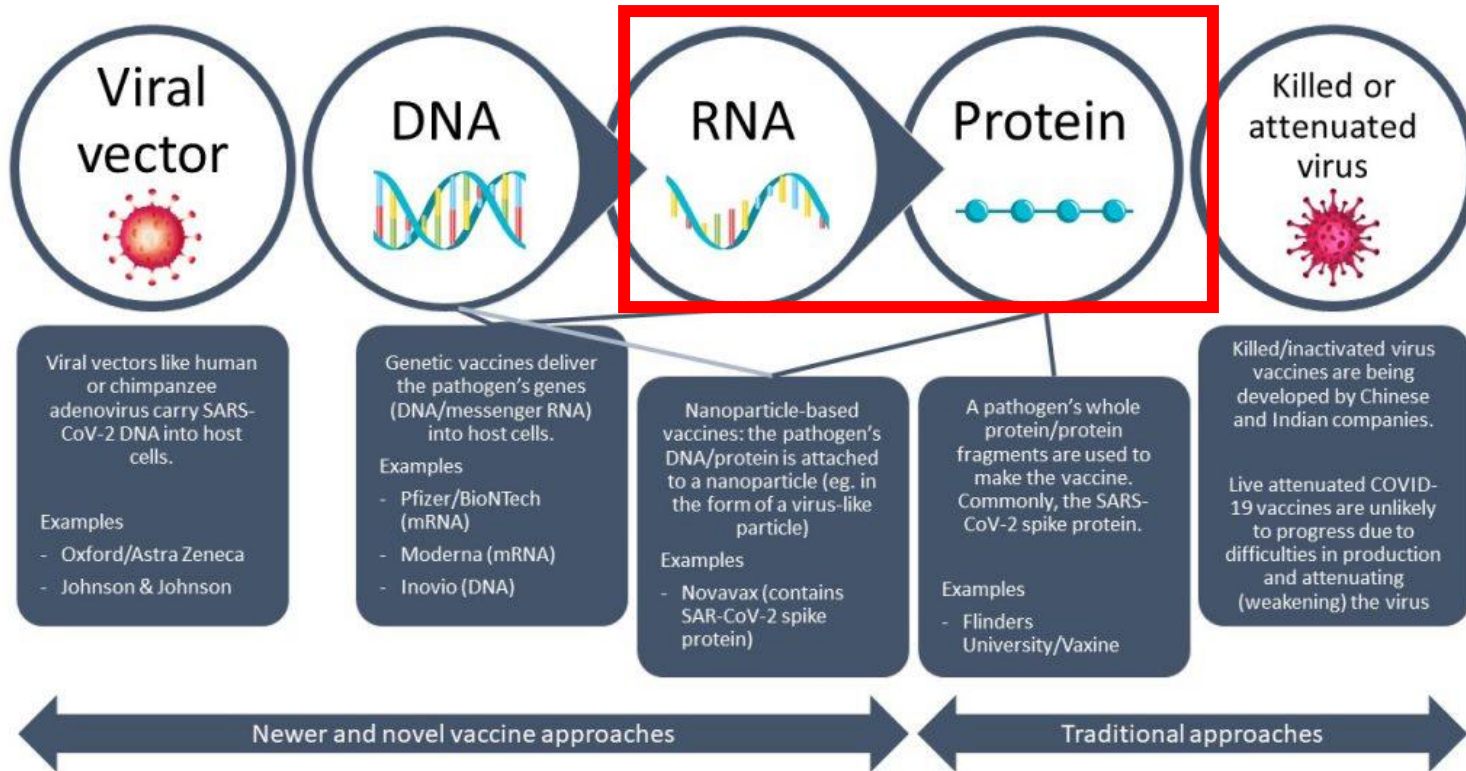
**CLC: 12 Actionable Fusions**

**Dragen/EasyFuse/Arriba: 1 Actionable Fusion**

# Peptide antigen and mRNA Vaccines for the Treatment & Prevention of Cancer



# Modern Approaches to Vaccination



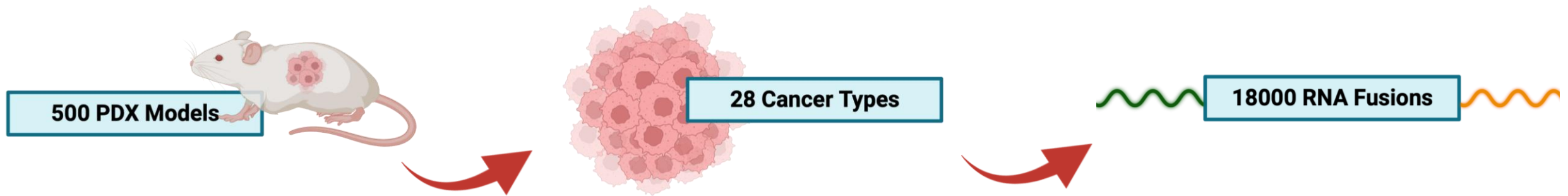
## Why Peptide Antigen Vaccines?

- Target very specific epitopes while minimizing the risks of allergic or autoimmune reactions

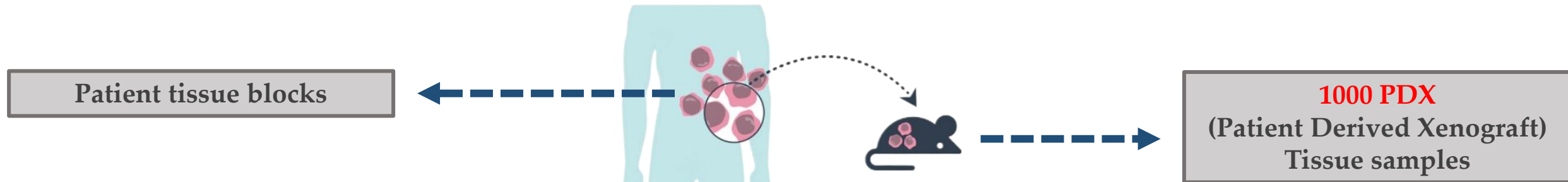
## Our focus?

- Extracting neoantigens from fusion transcripts from two separate genes
- The unique sequences at the fusion junctions form new open reading frames (ORFs)

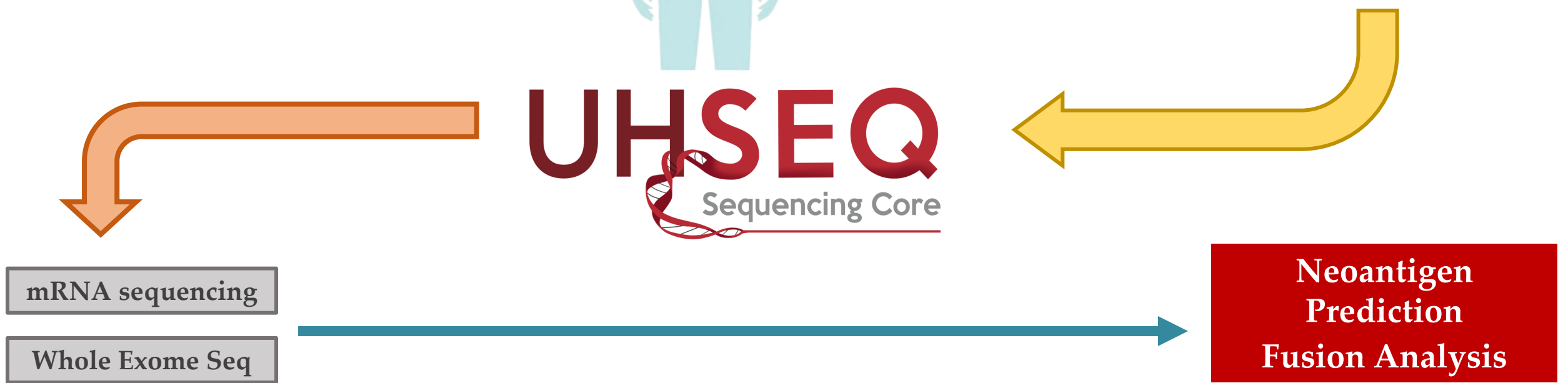




xenSTART

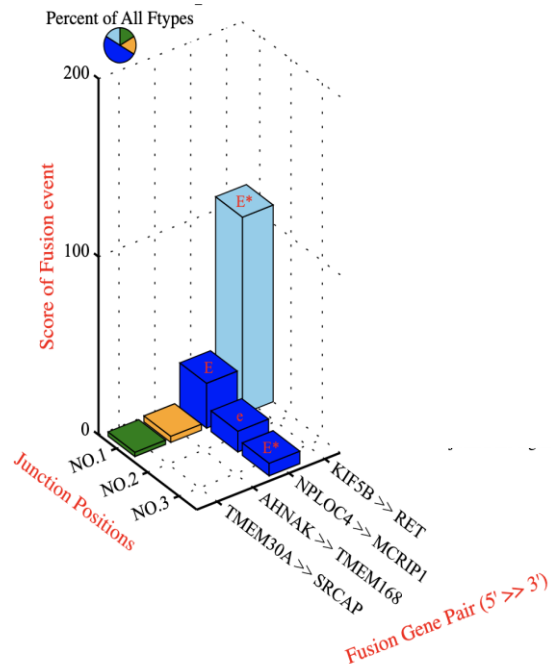


UHSEQ  
Sequencing Core



# RT-PCR and Sanger Validation of PDX Fusion KIF5B-RET

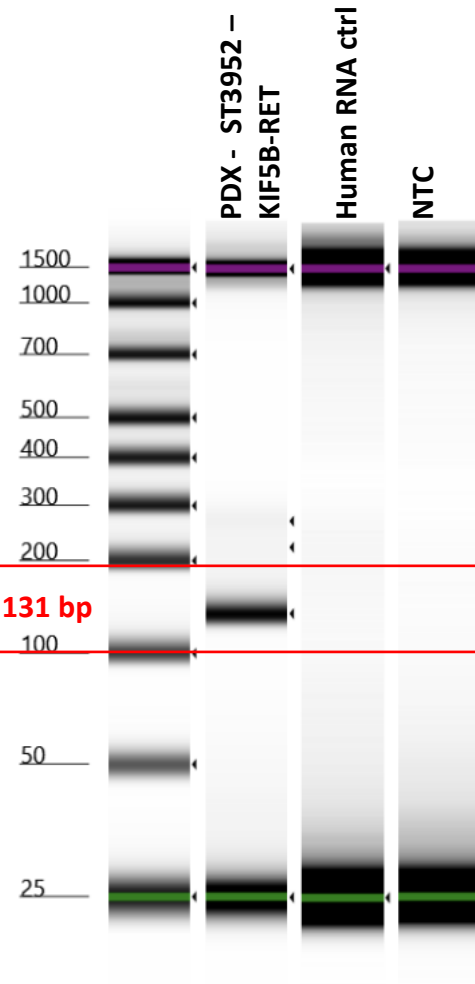
## In-silico Identification



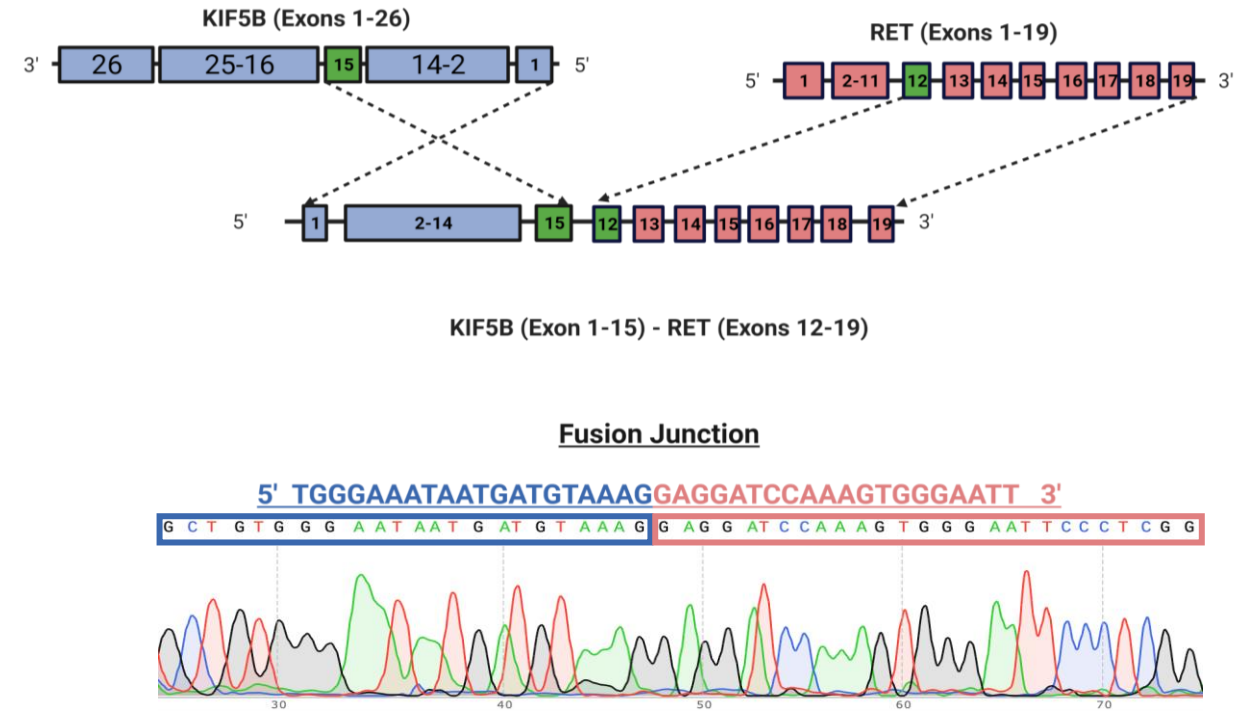
Pillar-Top-Text:  
\* inframe-shift  
e one-exon-edge  
E both-exon-edge

Pillar-Fill-Colour:  
INTERCHR-DS  
INTERCHR-SS  
INTRACHR-DS  
INTRACHR-SS-RGO

## RT-PCR

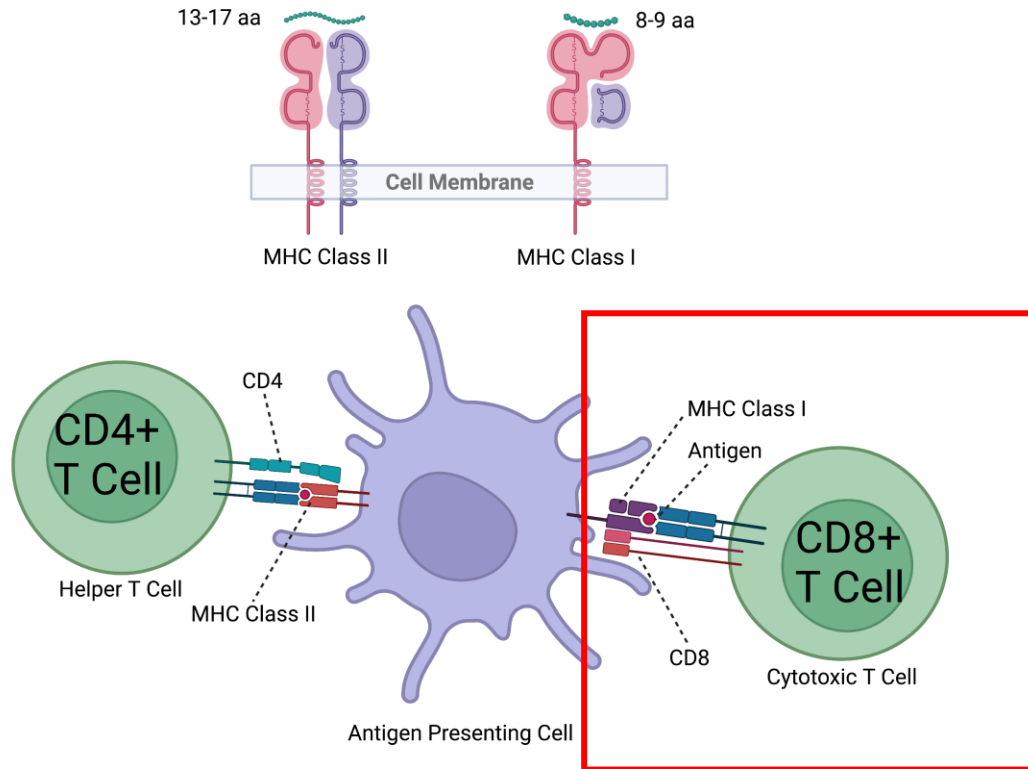


## Validation by Sanger Sequencing



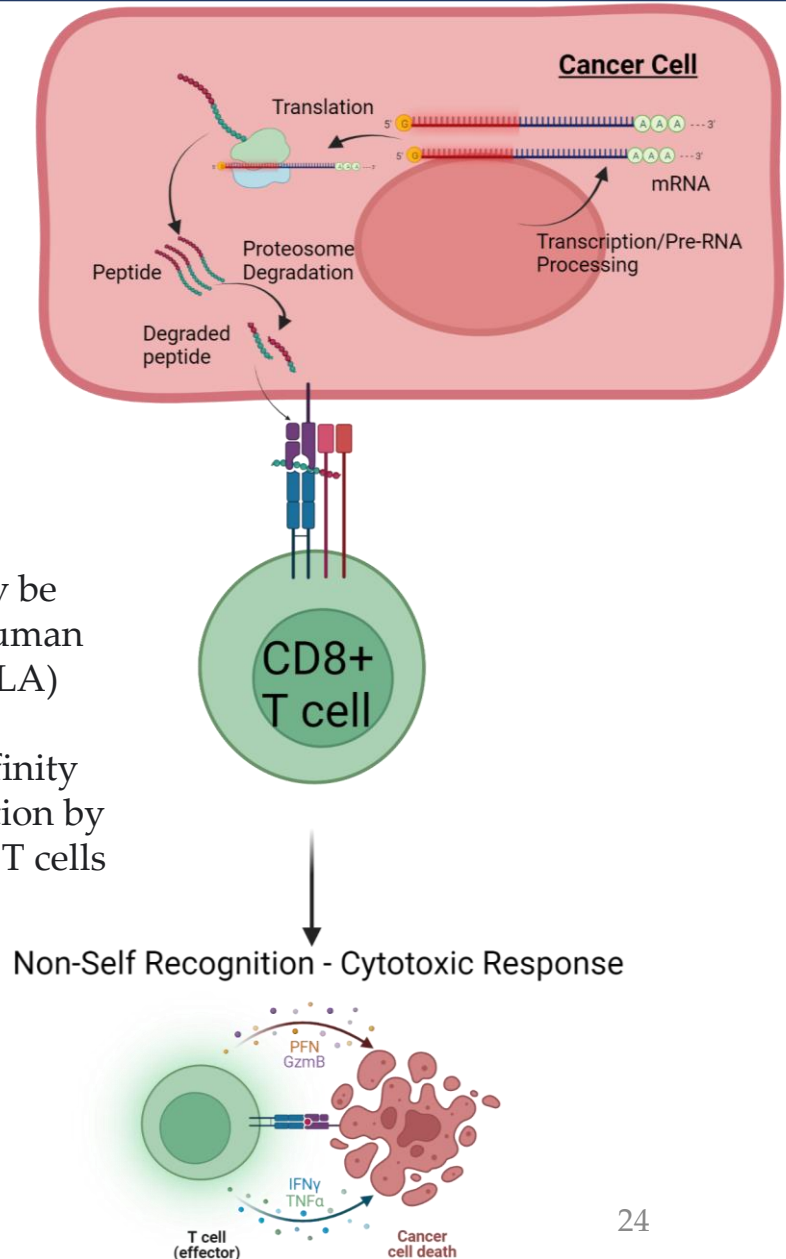


# Fusion Presentation to CD8+ and CD4+ Requires MHC-Class I & II



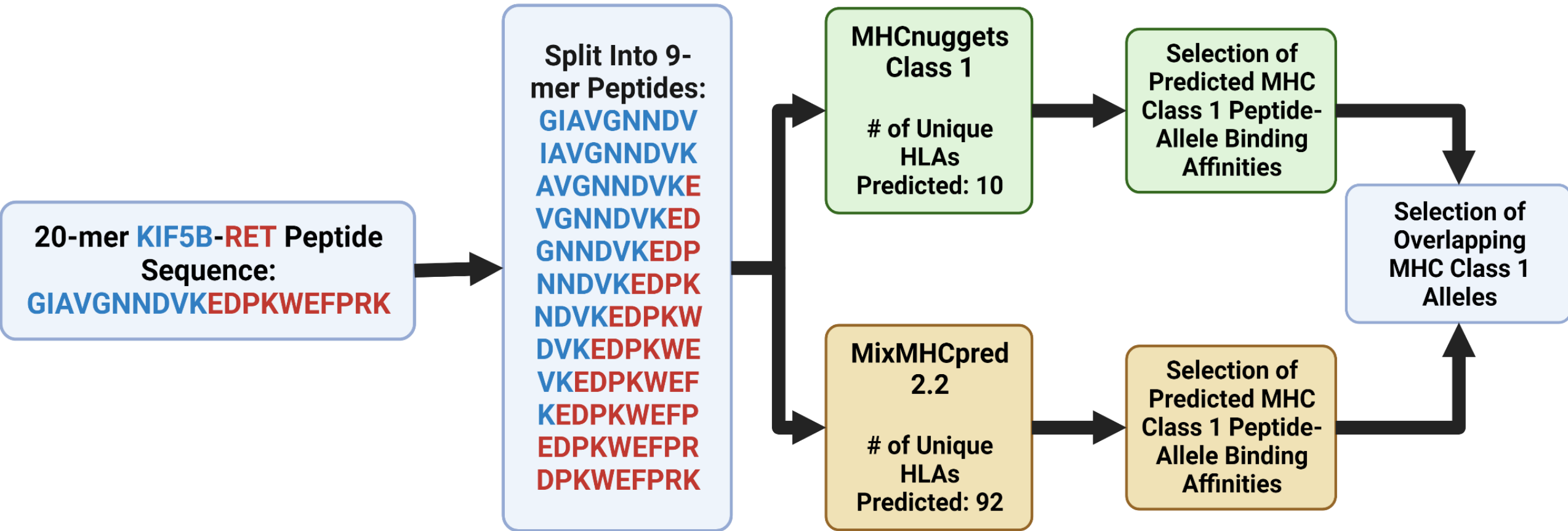
- CD8+ (cytotoxic) T cells directly kill cells presenting non-self epitopes.
- CD4+ (helper) T cells regulate the immune response to a particular antigen.

- Degraded proteins may be presented on MHC (Human Leukocyte Antigen - HLA) Class I molecules
  - Dependent on affinity
  - Non-self recognition by circulating CD8+ T cells





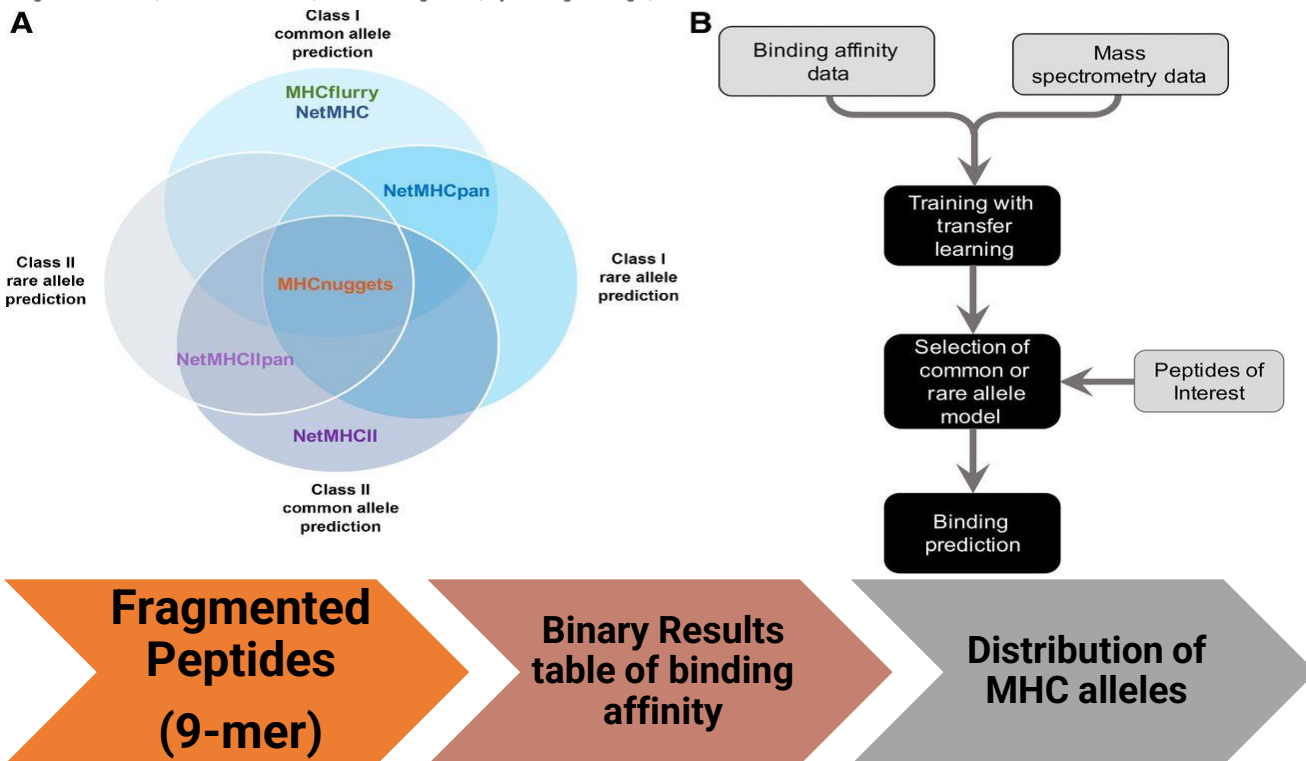
# Neopeptide Affinity Prediction Pipeline



# Neopeptide Affinity Prediction Pipeline

## High-Throughput Prediction of MHC Class I and II Neoantigens with MHCnuggets

Xiaoshan M. Shao, Rohit Bhattacharya, Justin Huang, I.K. Ashok Sivakumar, Collin Tokheim, Lily Zheng, Dylan Hirsch, Benjamin Kaminow, Ashton Omdahl, Maria Bonsack, Angelika B. Riemer, Victor E. Velculescu, Valsamo Anagnostou, Kymberleigh A. Pagel, and Rachel Karchin



- Deep-learning models
- Transfer-learning protocol
  - Improves predictive power by circumventing issues due to variability of peptide length and the lack of data concerning rare HLA alleles.

## GfellerLab/ MixMHCpred

HLA-I ligand predictor



1

Contributor



0

Issues



20

Stars



7

Forks



- Uses sequence-scoring functions as basis of predictions.
  - Calculates Position-Specific Scoring Matrix (PSSM) of all the inputted HLA alleles.
  - Outputs %Rank value indicative of HLA allotype's binding affinity to a specified peptide.
  - Sensitivity (Sn), Specificity (Sp), Accuracy (Acc)
- Cutoff value of 10.00 for %Rank values was considered a strong binder
  - HLA alleles that fell within this cutoff value often had IC50 values less than 500nM when compared to results from MHCnuggets.

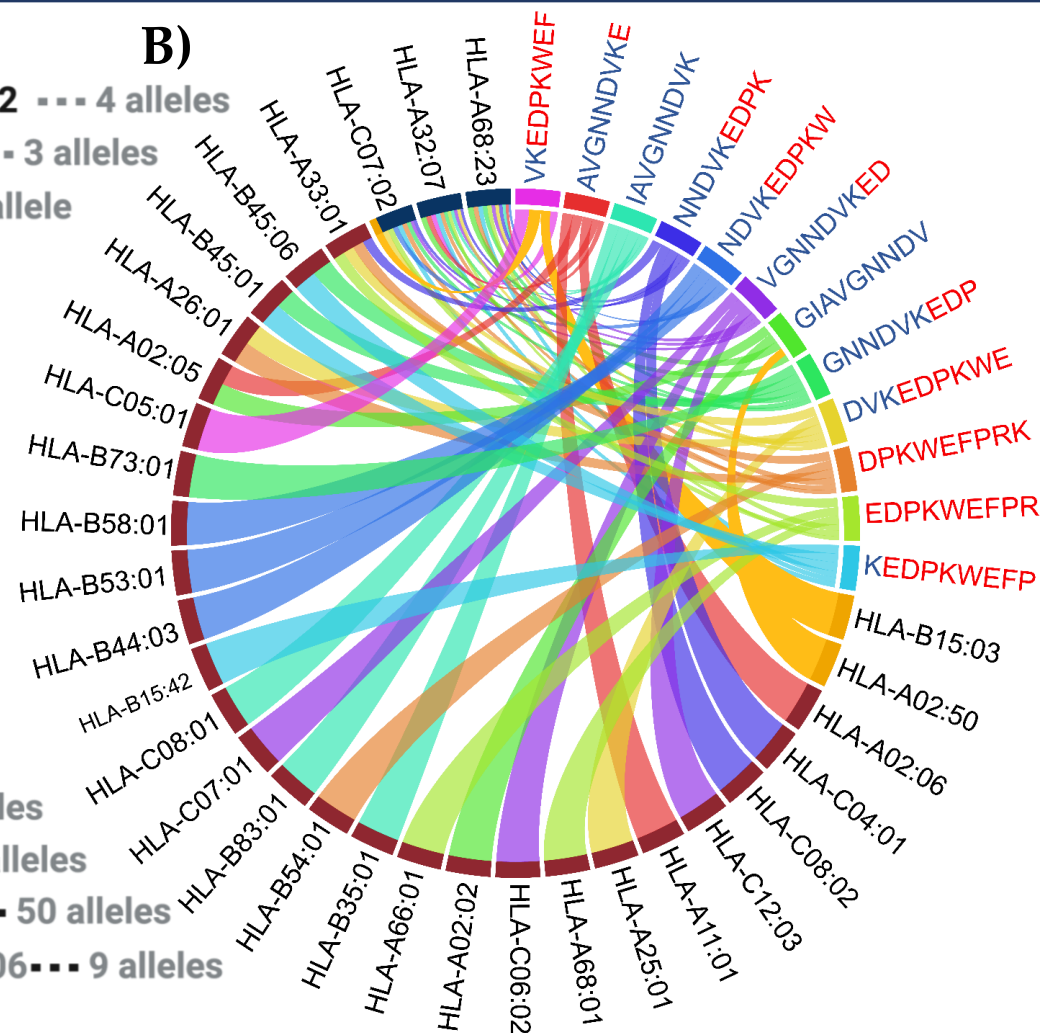
# Neopeptide Affinity Predictions

A)



A) Binding of junction peptides to MHC Class 1 alleles. Lines in **grey** represent calls made by MHCNuggets. Lines in **black** represent calls made by MixMHCPred.

B)



B) Circos plot of neoantigen binding. **Red** and **blue** origins denote HLAs called by MixMHCPred and MHCNuggets, respectively. **Gold** chords denote overlapping calls.

# HLA-C07:02 Shows Strong Binding to KIF5B-RET Neopeptides

HLA Allele	GIAGNNDV	IAVGNDVK	AVGNNDVKE	VGNDVKE	GNDVKE	NNDVKE	NDVKE	DKVKE	VKE	KEDVKE	EDVKE	EDVKE
HLA-A02:02	2.2											
HLA-A02:05	1.3		8.9									
HLA-A02:06			9.5									
HLA-A02:50	15/2.17142											
HLA-A11:01			4.8									
HLA-A25:01								5.7				
HLA-A26:01								5.9				3.3
HLA-A32:07	38.7	229.8	43.3	227.9	154.1	385.3		219.0	60.5	55.2	174.5	115.0
HLA-A33:01						8.8		2.0			0.8	2.0
HLA-A66:01											1.3	
HLA-A68:01											4.3	
HLA-A68:23	13.3	95.7	154.3	76.2	37.0	84.4	489.0	76.6		29.2	36.4	26.8
HLA-B15:03									29.33/0.251			
HLA-B15:42										0.1		
HLA-B35:01		1.9										
HLA-B44:03							1.0					
HLA-B45:01					6.9					0.2		
HLA-B45:06					6.9					0.2		
HLA-B53:01							0.6					
HLA-B54:01												2.8
HLA-B58:01							0.5					
HLA-B73:01					0.8							
HLA-B83:01		1.9										
HLA-C04:01						9.4						
HLA-C05:01									0.5			
HLA-C06:02				4.5								
HLA-C07:01				2.5								
HLA-C07:02				310.6	102.8	295.0	48.2	39.1	8.82/0.211	55.6	39.8	57.0
HLA-C08:01		1.8										
HLA-C08:02						7.5						
HLA-C12:03				7.7								

**Table 1:** Neoantigen affinity prediction table. Red and blue cells denote HLAs called by MixMHCpred and MHCNuggets, respectively. Gold cells denote overlapping calls.



# Structure Based Binding Affinities and Peptide Cross-reactivity



Geometry prediction for peptide-HLAs

Binding energy prediction for peptides

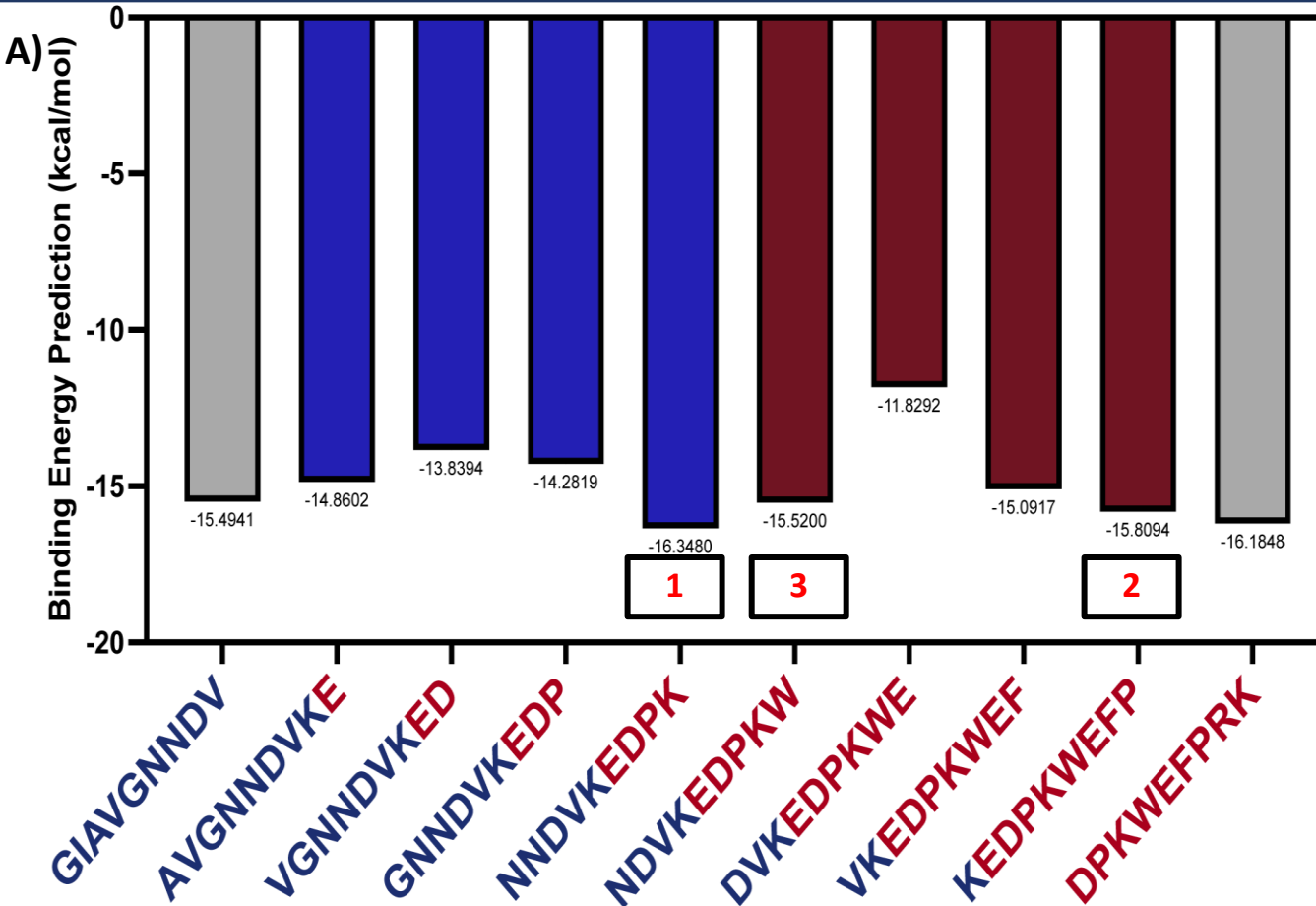
Virtual screening of HLA binders

Customizable workflows for analysis

## Considerations Taken

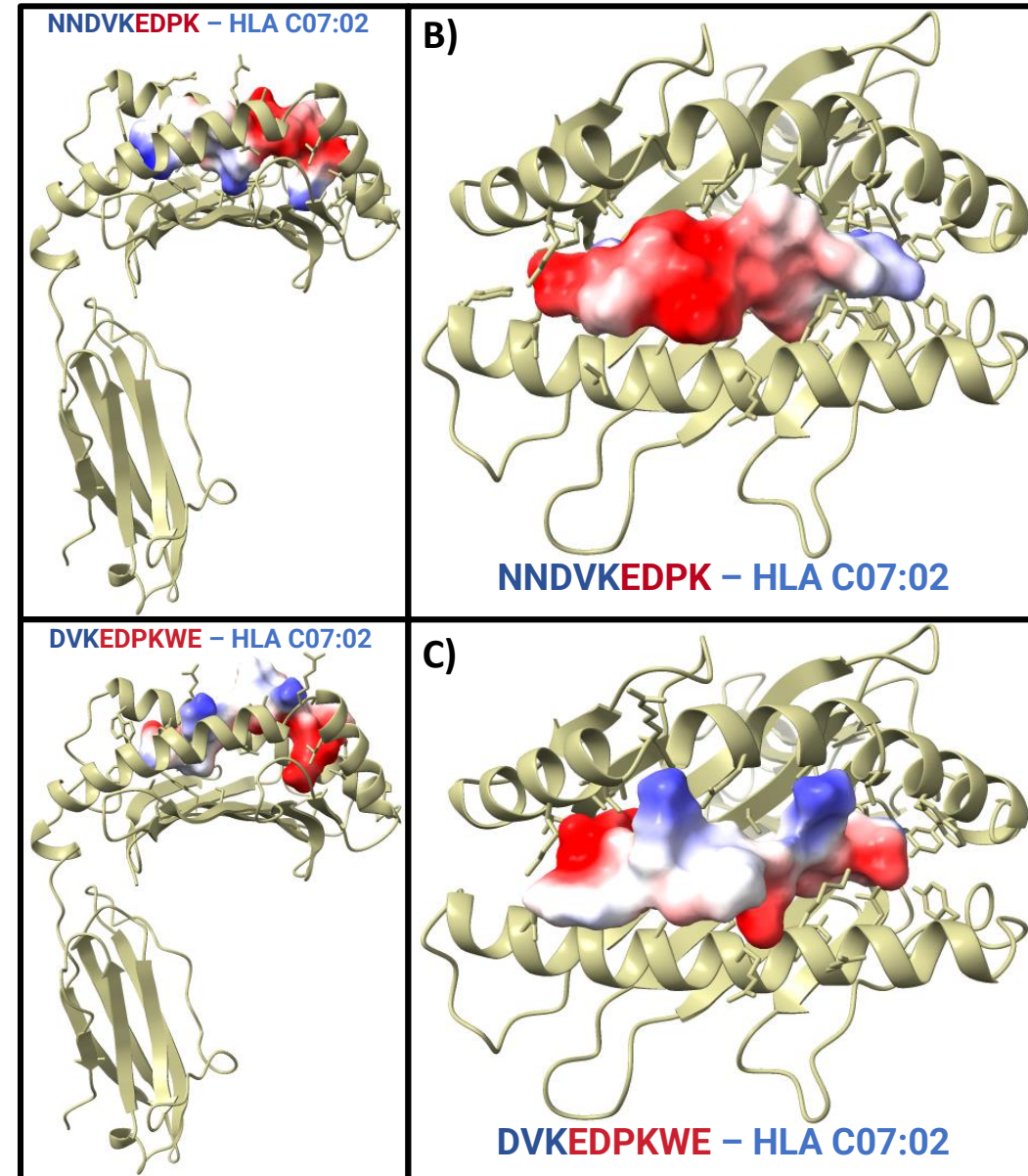
- APE-GEN used to model 8 junction and 2 wild type peptides to the HLA-C\*07:02 receptor.
- HLA-C\*07:02 crystal structure 5VGE sourced from Protein Data Bank (PDB).
- R package Bio3d used to remove excess molecules and verify no chain breaks or gaps.
- Binding energies calculated with rigid structure.

# Structure Based Binding Affinity Predictions



Binding Strength

Binding Energy



**HLA-Arena Binding Energy Predictions and HLA-Peptide Docking.** A) Binding energy predictions for junction peptides and two WT peptides (grey). Bars colored by majority contribution of amino acids in blue and red for KIF5B and RET respectively. B) and C) Peptides with highest and lowest average binding energy prediction values across three replicates. Peptides are docked in HLA-C07:02 and are colored by their electrostatic potential.

In Collaboration with: Jaila Lewis and Martiela Vaz de Freitas (Antunes Lab)

# Structure Based Binding Affinities and Peptide Cross-reactivity



Geometry prediction for peptide-HLAs

Binding energy prediction for peptides

Virtual screening of HLA binders

Customizable workflows for analysis

## Considerations Taken

- APE-GEN used to model 8 junction and 2 wild type peptides to the HLA-C\*07:02 receptor.
- HLA-C\*07:02 crystal structure 5VGE sourced from Protein Data Bank (PDB).
- R package Bio3d used to remove excess molecules and verify no chain breaks or gaps.
- Binding energies calculated with rigid



Maps to Immunopeptidomic Data

Identifies Self Derived Peptides

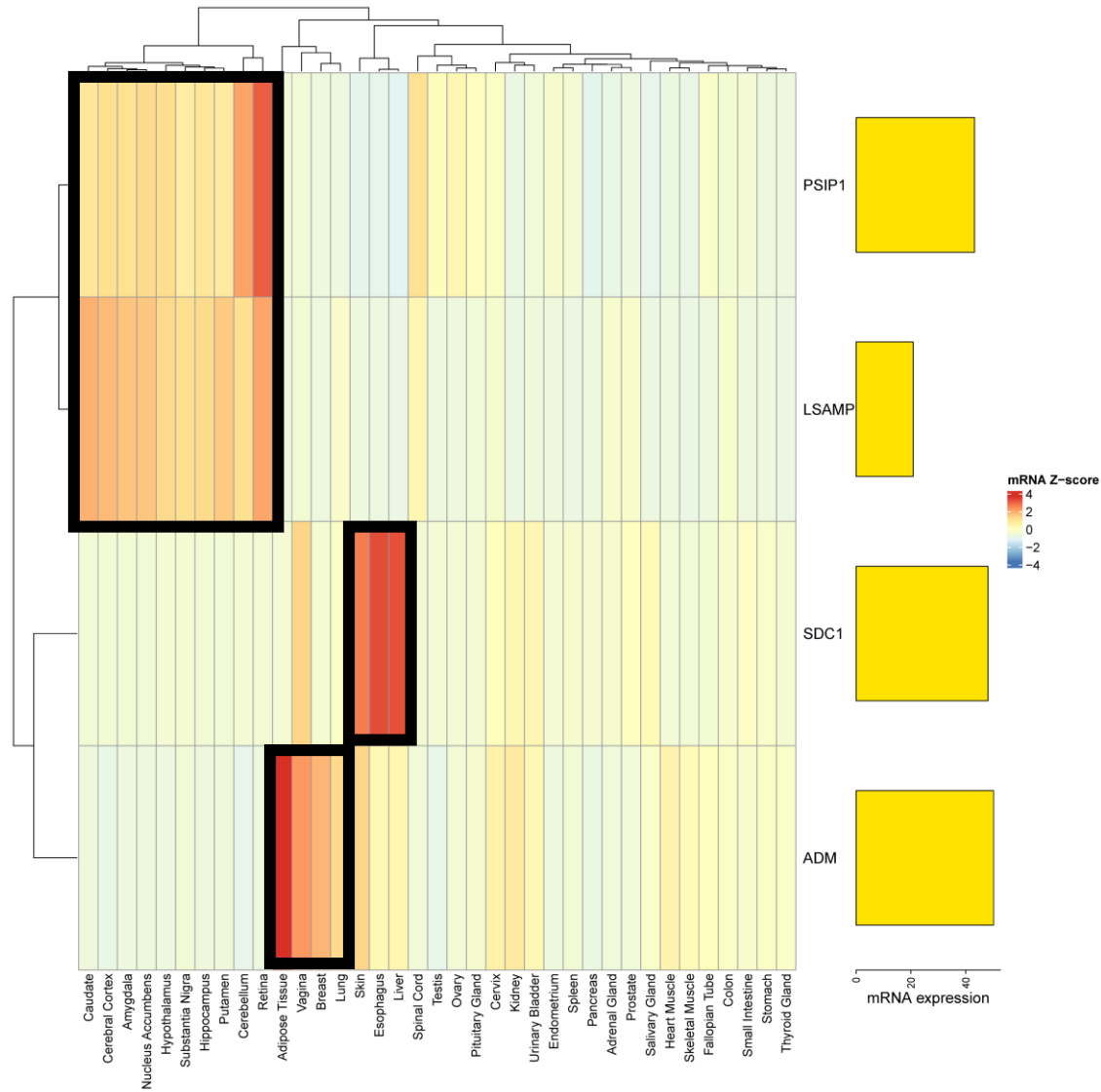
Tissue Specific mRNA Expression

## Considerations Taken

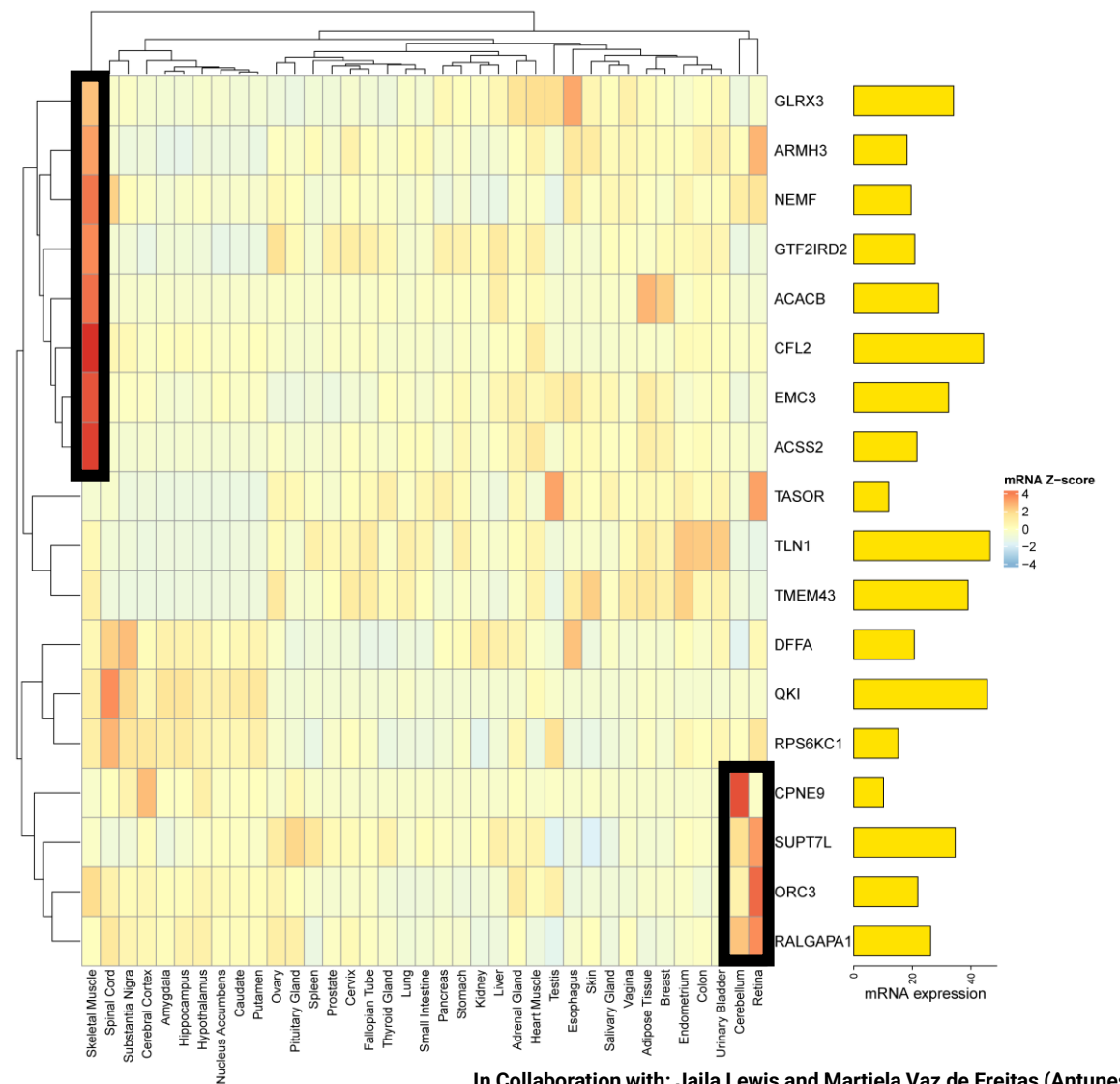
- 9-mers were limited to the HLA-C\*07:02 allele to identify all potential self-derived peptides that may yield T-cell cross-reactivity.
- Default p-value cutoff for cross reactivity  $p < 0.005$ 
  - Increased to  $<0.01$  and  $<0.05$  for this dataset

# Assessment of T-Cell Cross Reactivity

NNDVKEDPK

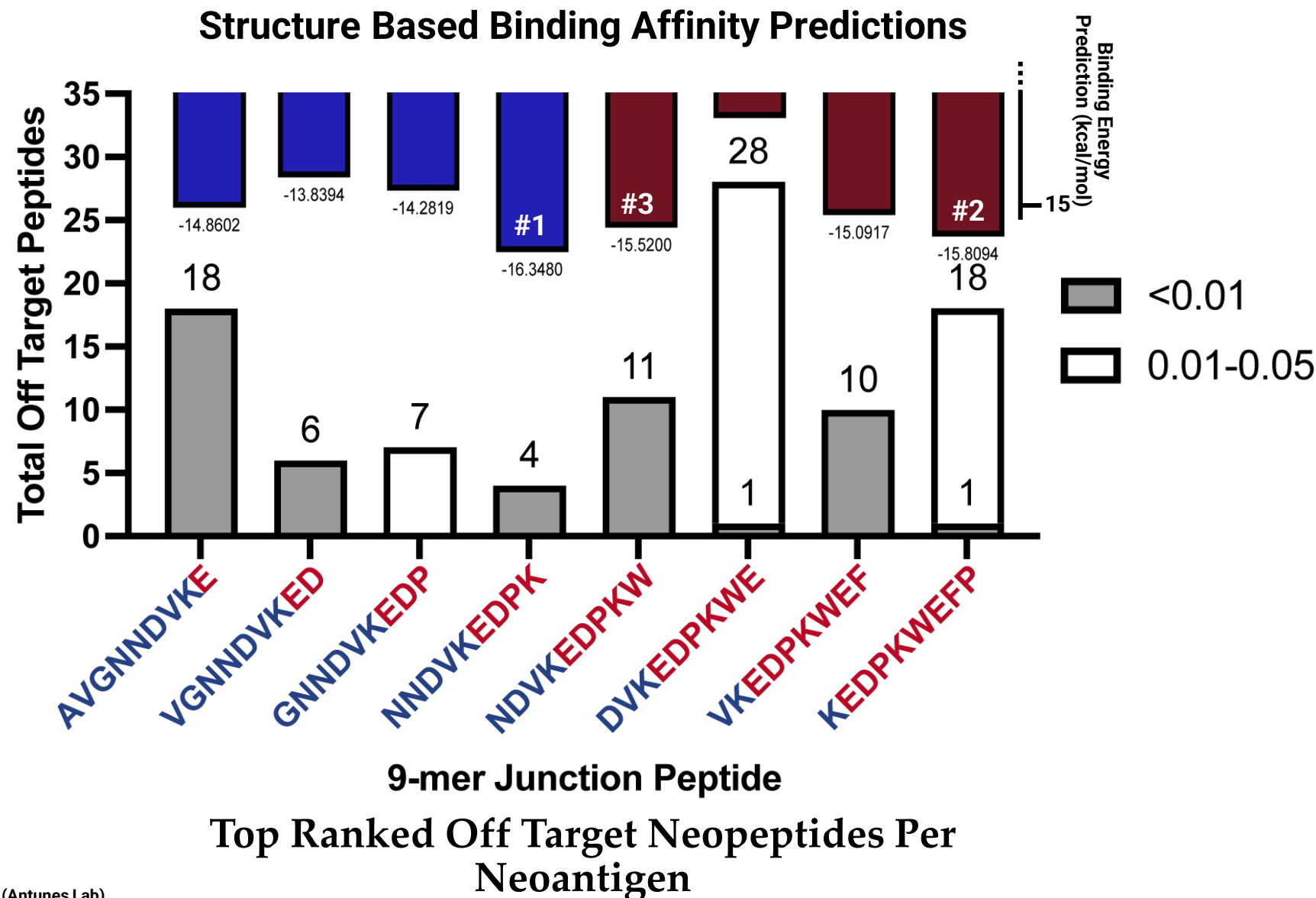


KEDPKWEFP

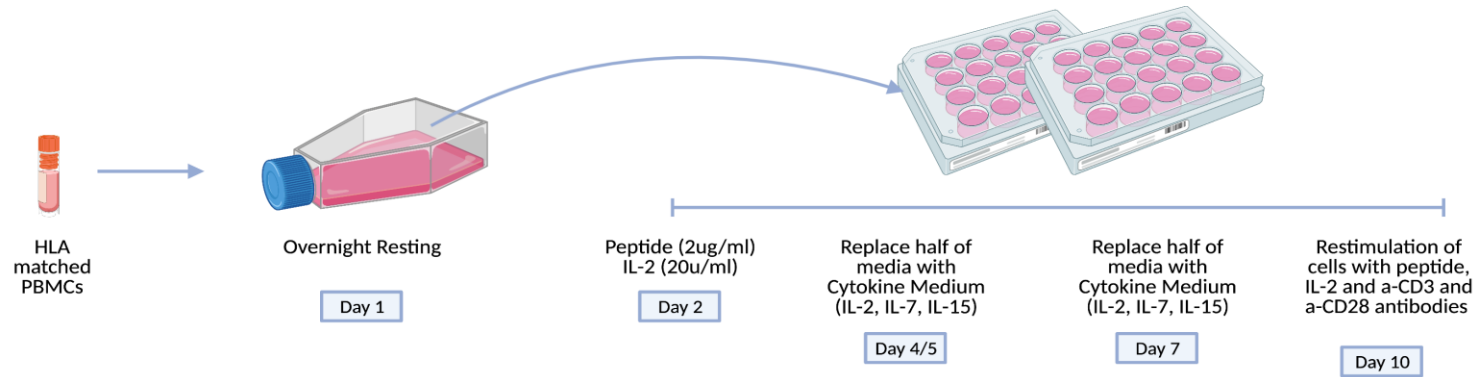




# Crossdome Assessment of T-Cell Cross Reactivity

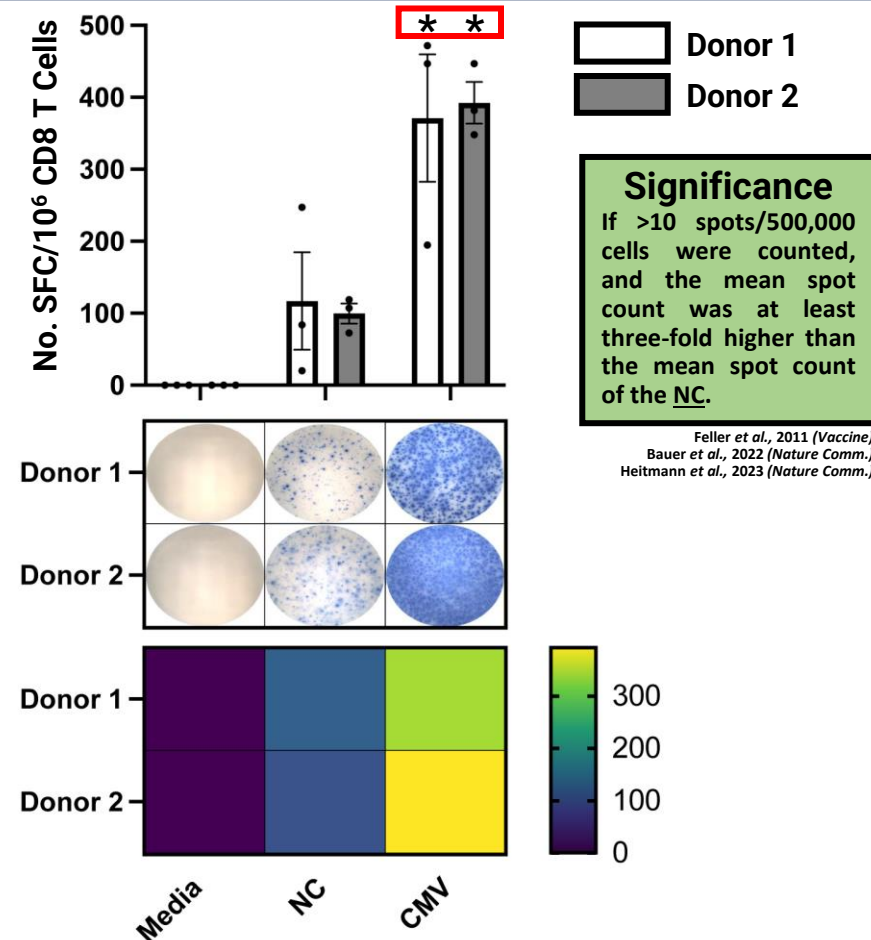
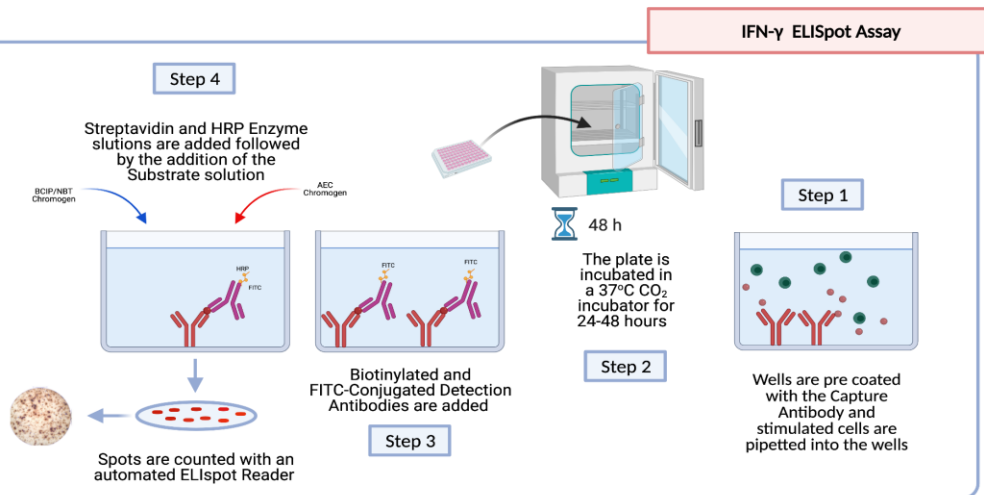


# Enzyme Linked Immunosorbent Spot Assay (ELISpot)

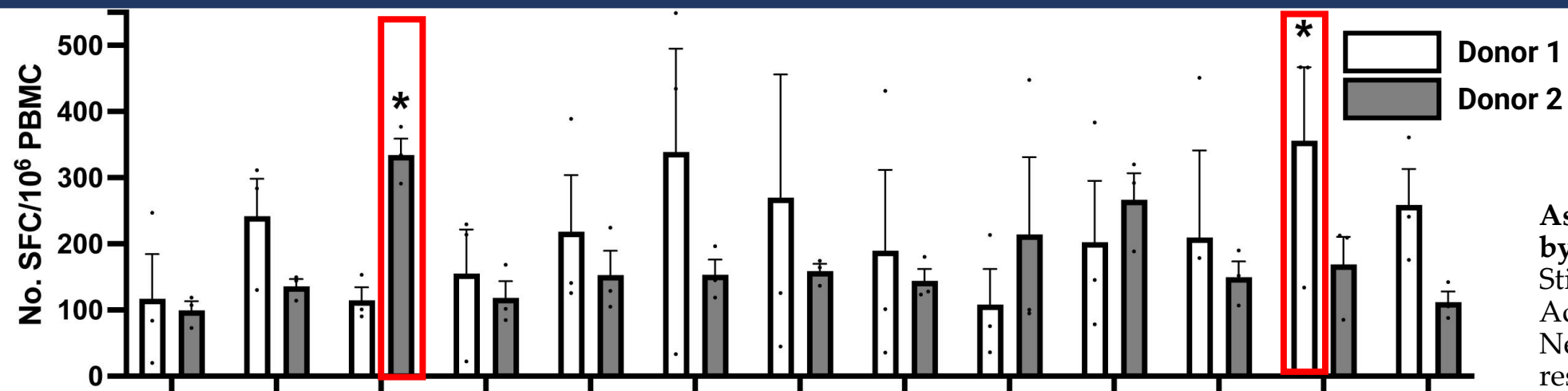


Day 12/13/14  
Negative Selection of CD8 cells from Activated PBMCs

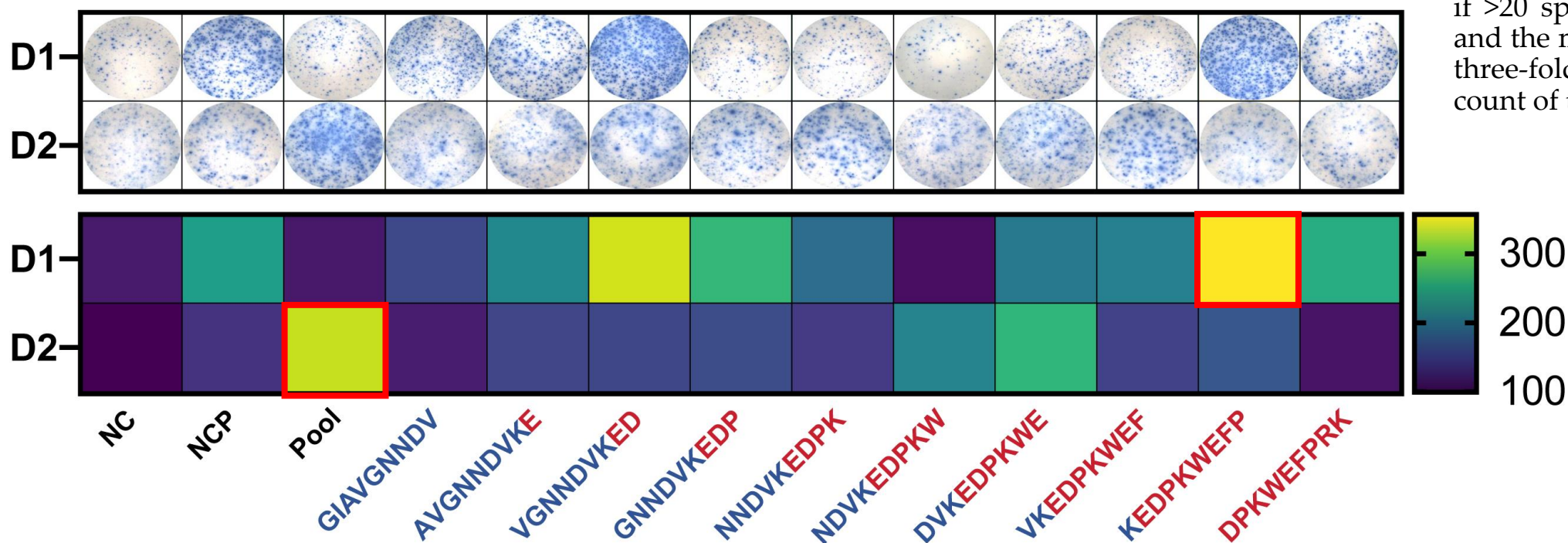
IFN-γ Colour ELISpot Assay using the CD8 cells



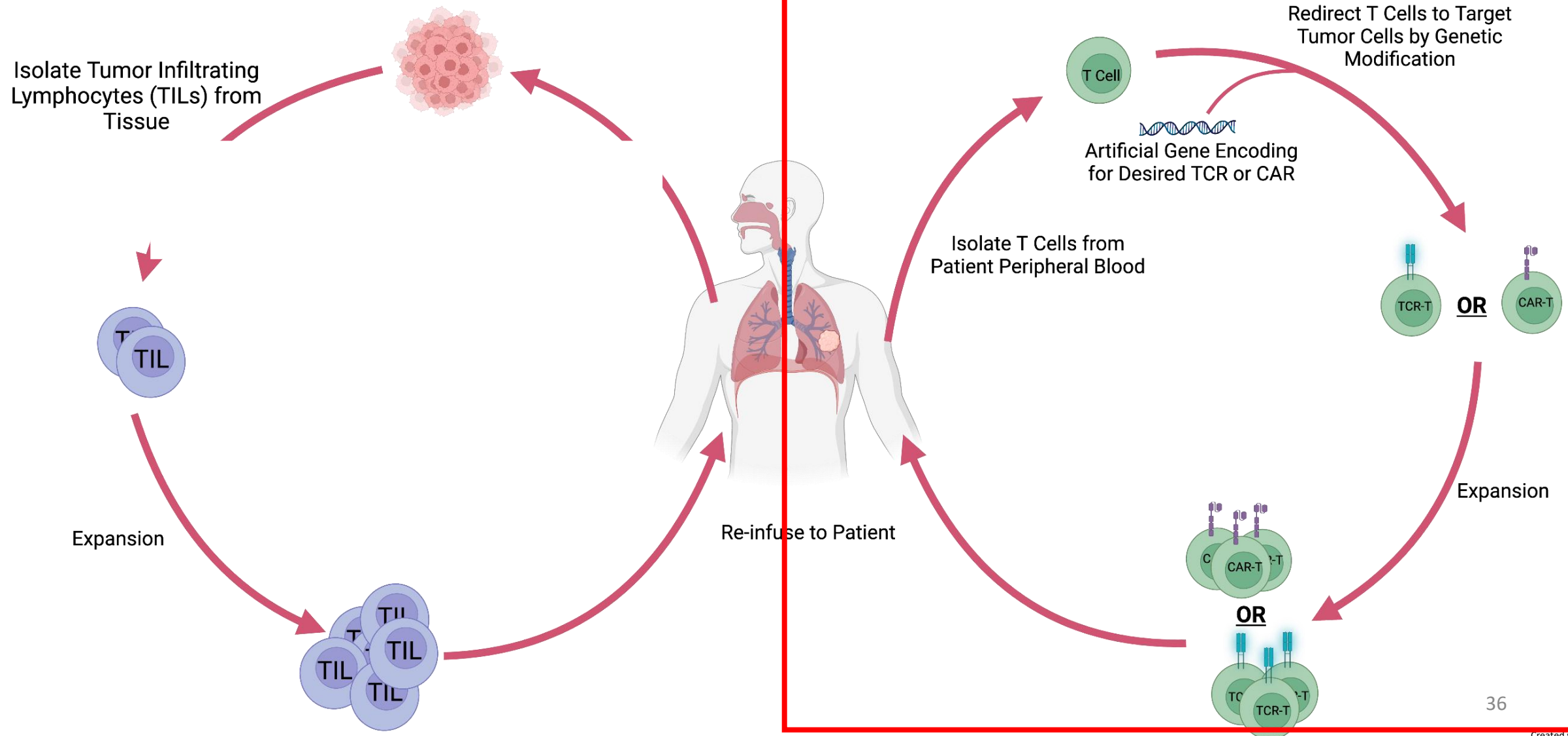
# *In-vitro* Peptide Stimulation Reveals Responses Varying By Donor



**Assessing CD8+ T Cell Stimulation by IFN $\gamma$  ELISpot.** NC: Non-Stimulated Control, NCP: Non-Activation Control Peptide, NJP: Neoantigen Junction Pool. T cell responses were considered positive if >20 spots/1M cells were counted, and the mean spot count was at least three-fold higher than the mean spot count of the NC.

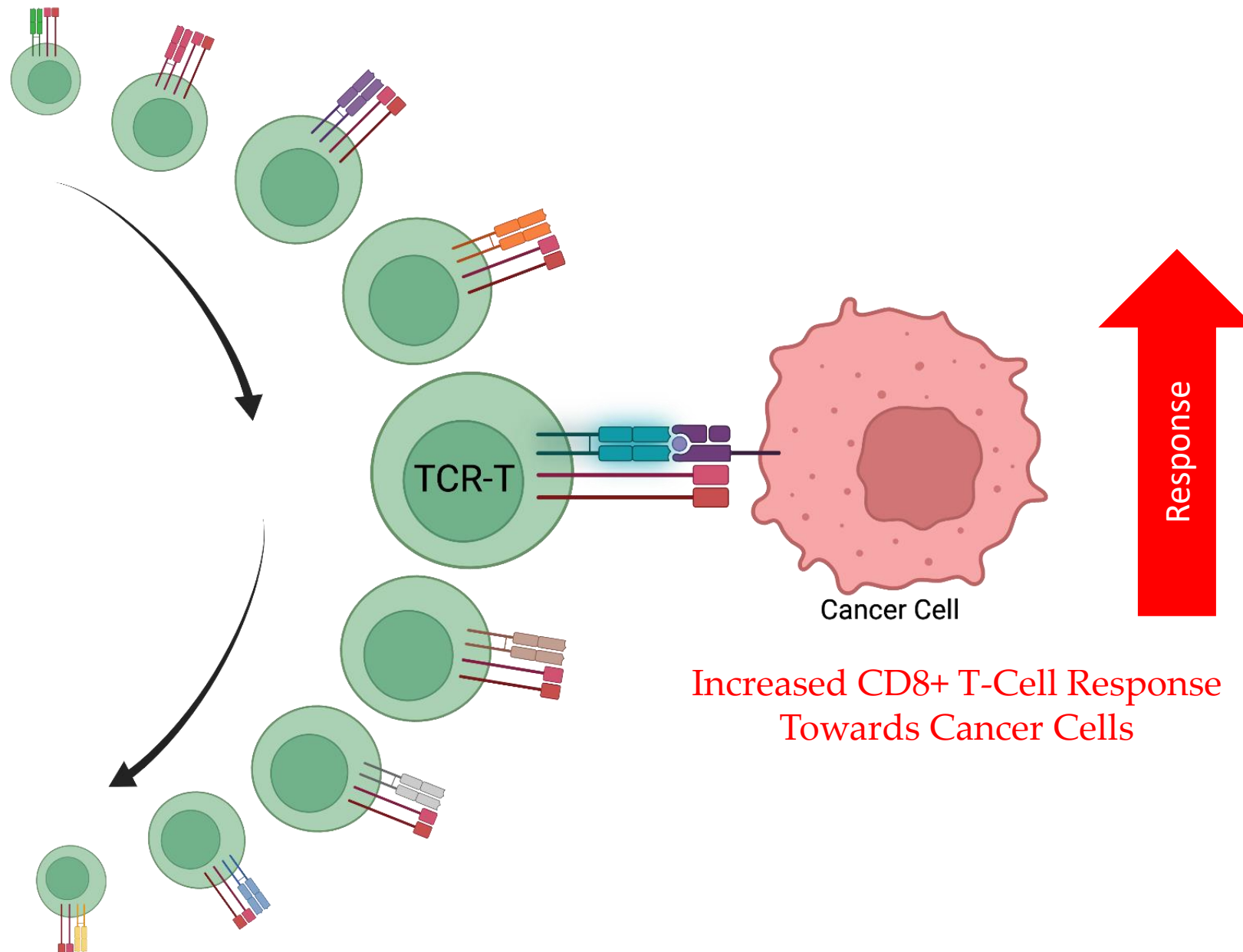


# Adoptive Cell Immune Therapies

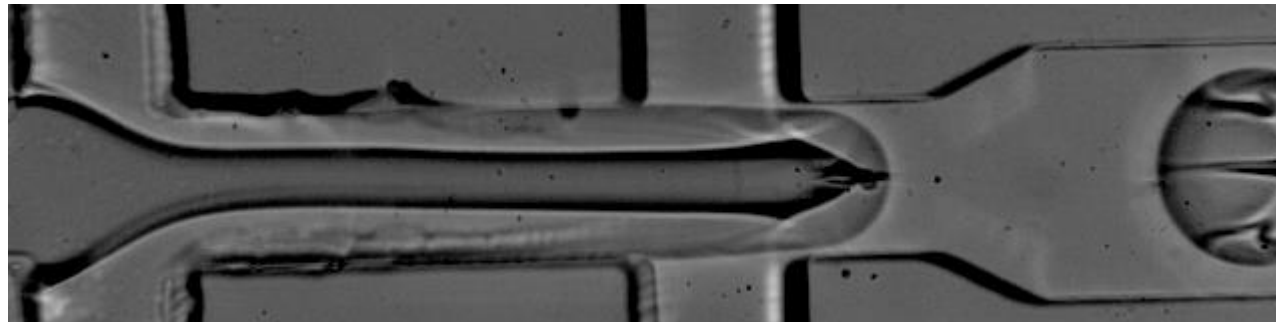
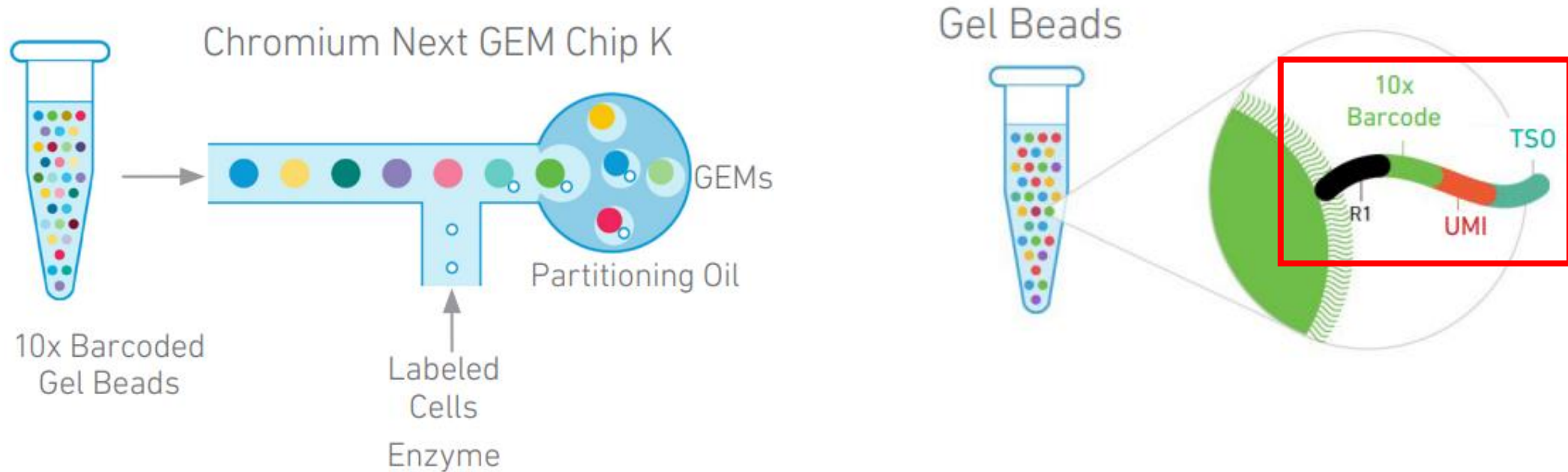




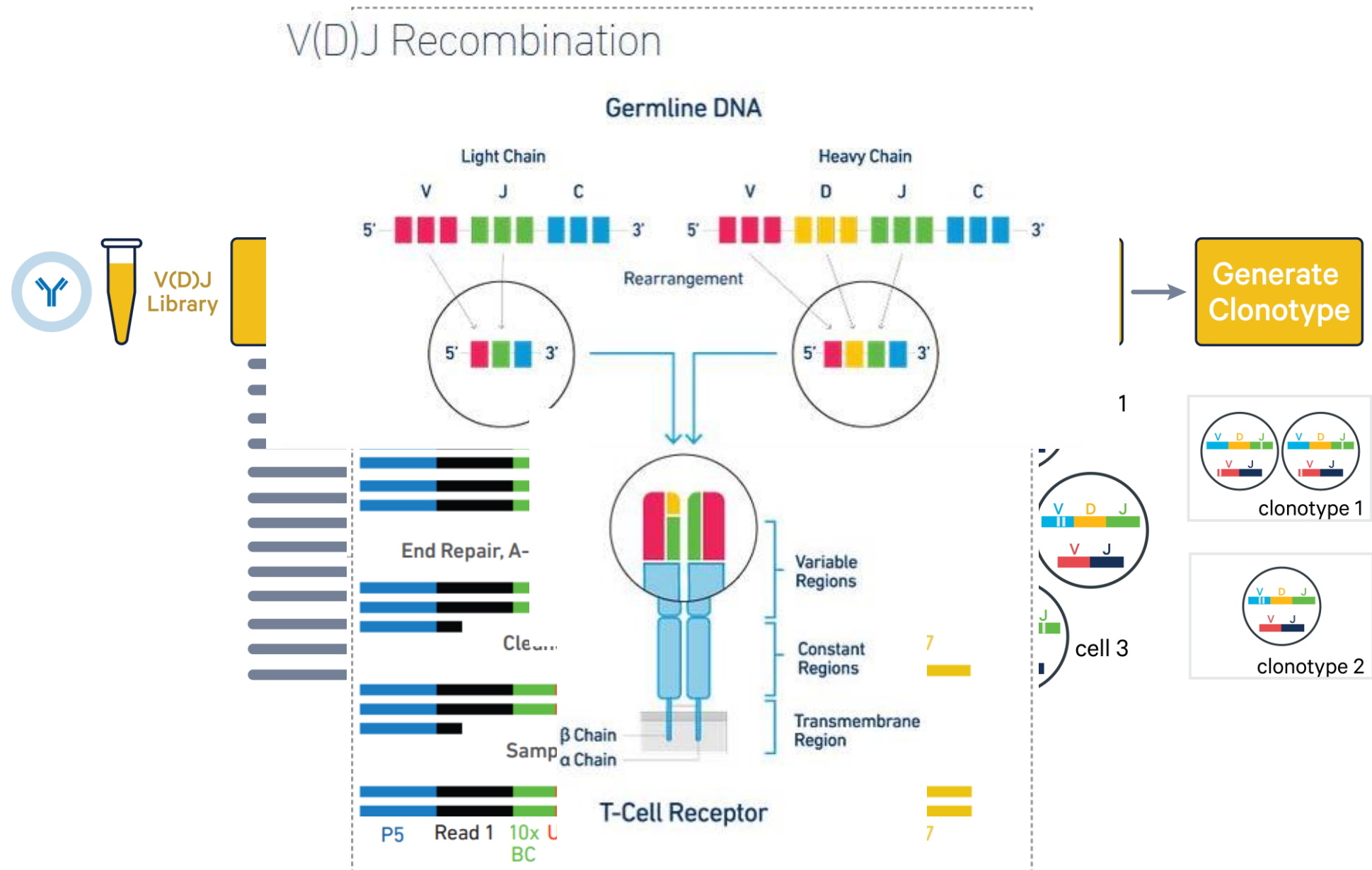
# Identification of Expanded TCR Clonotypes in Peptide Stimulated T-Cells



# 10X Genomics Single Cell 5' Gene Expression and Immune Profiling

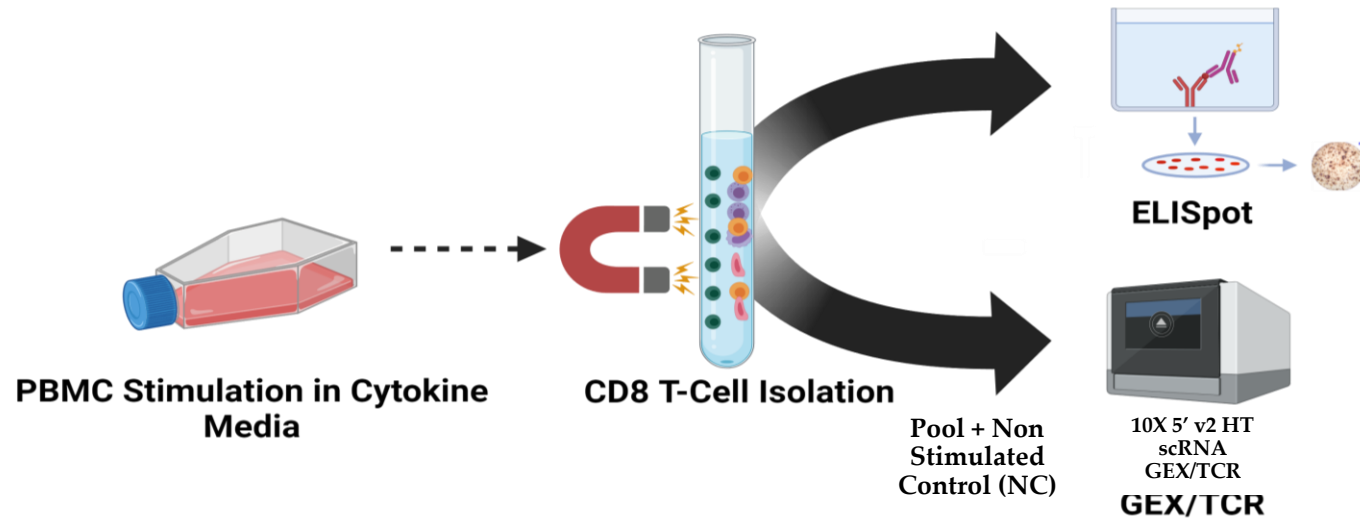


# VDJ Biology - Variability, (Diversity), Joining



# Single Cell Immune Profiling: Sample Prep/Sequencing/Data QC

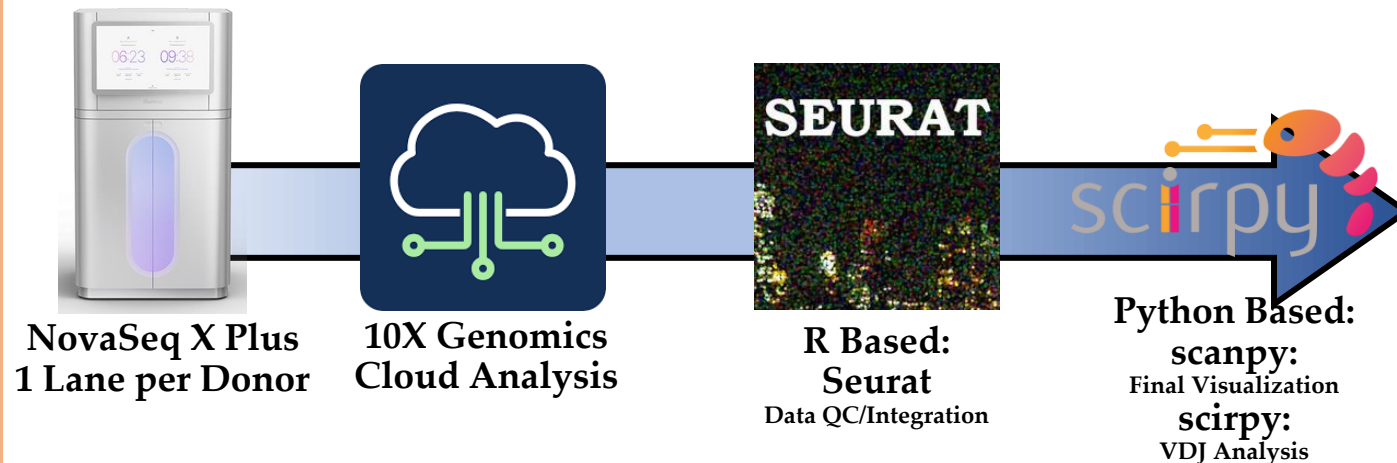
## Sample Prep



### Considerations Taken

- Cell Viability after CD8 T-Cell Isolation
  - 65-80% Live Cells
- Time between isolation and microfluidics chip loading
  - ~30 minutes
- Cells targeted
  - Donor 1: Lower cell counts elicited full sample loading
  - Donor 2: 12,000 cells

## Sequencing + Data Analysis

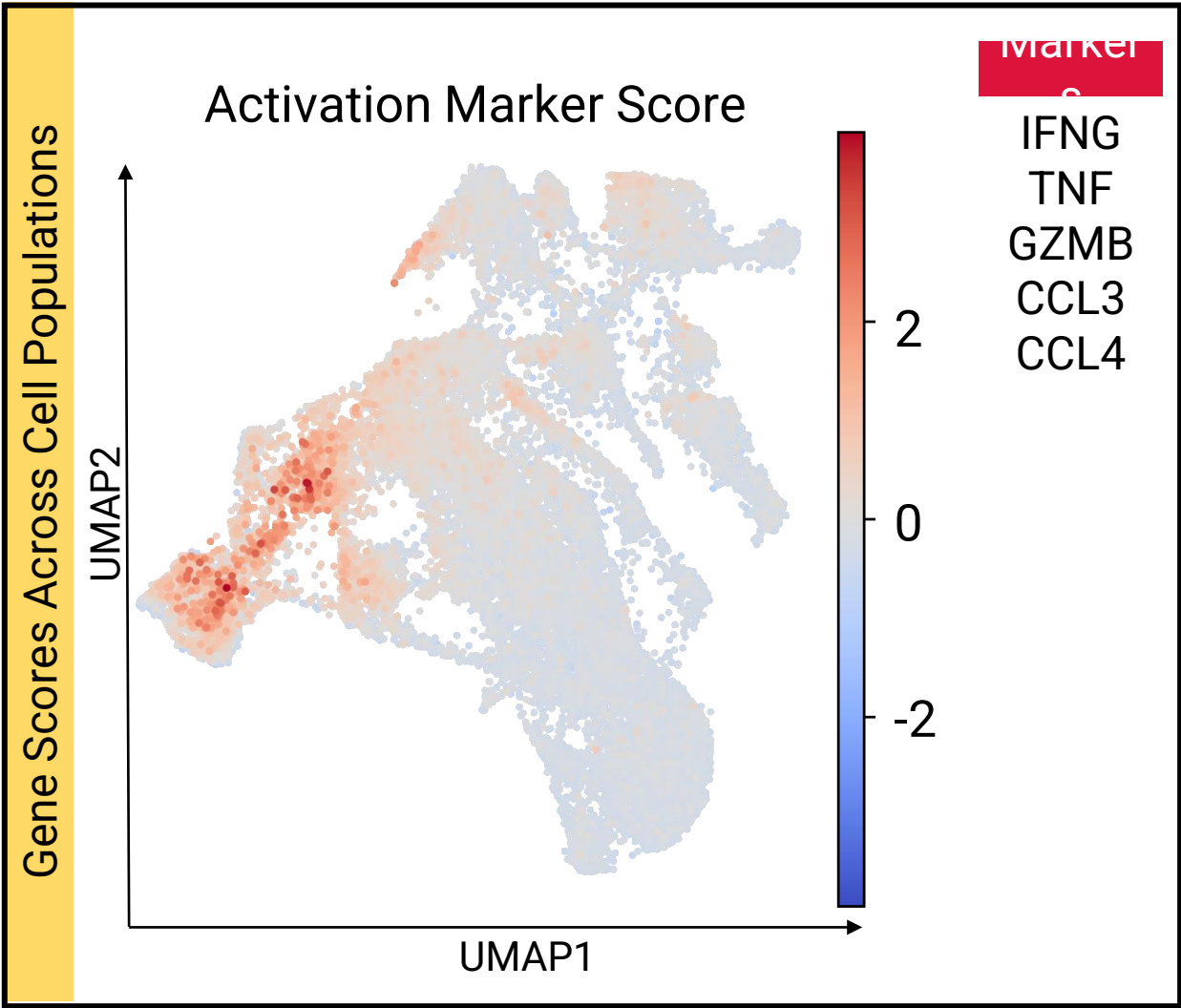
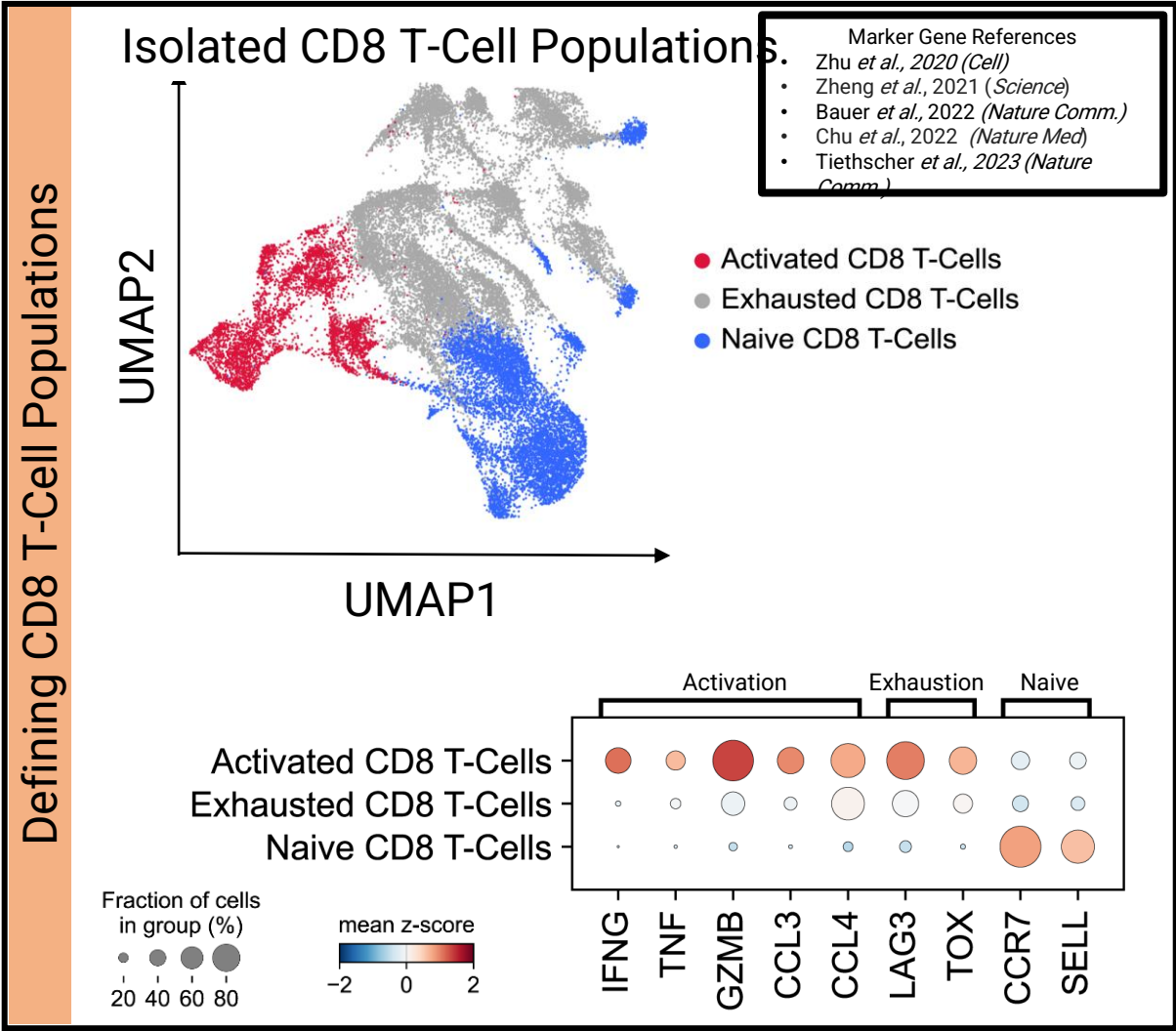


### Considerations Taken

- NGS libraries pooled 80/20:GEX/TCR
- Cell Ranger Count/V(D)J v7.1.0
- Doublets were removed using DoubletFinder and doublet rates specified by 10X 5' HT protocol (UG:CG000423.RevC).
- Cells with high (>5%) mitochondrial counts or low features counts (<500) were removed.
- Seurat objects were converted to scanpy .h5ad files by converting metadata components into python (scirpy) inputs.



# Activated, Exhausted, and Naïve T-Cell Populations Called Based on Canonical Markers

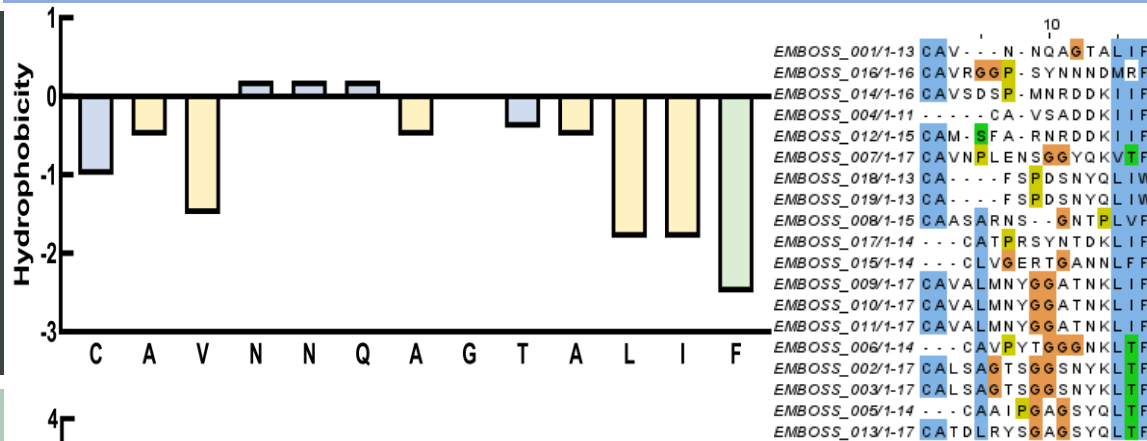


# Characterization of Top Expanded TCR Clonotypes

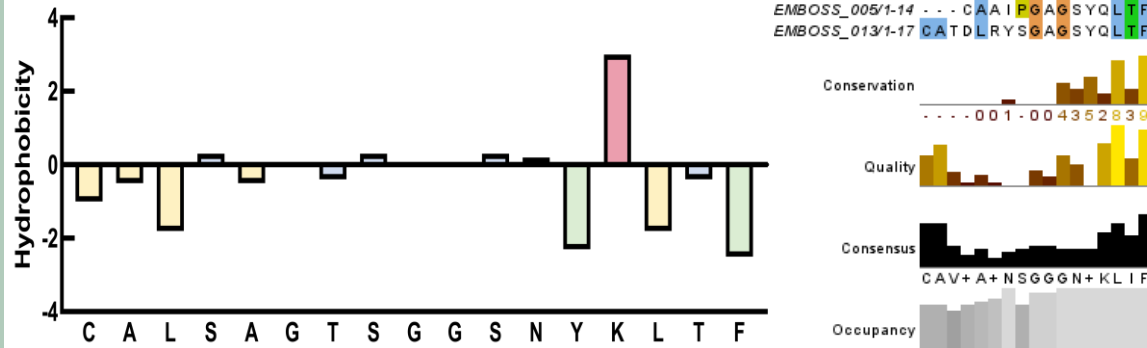
## Amino Acid View of Top Expanded TCRs Per Donor

### CDR3-α Chain

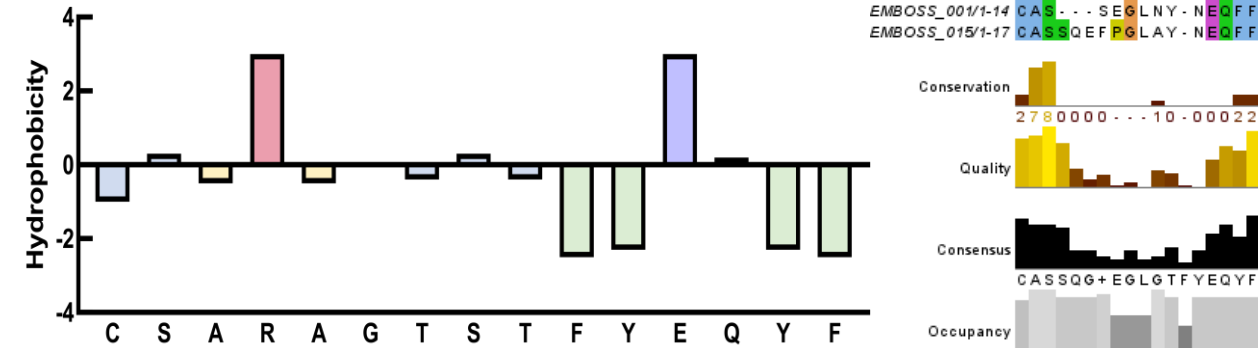
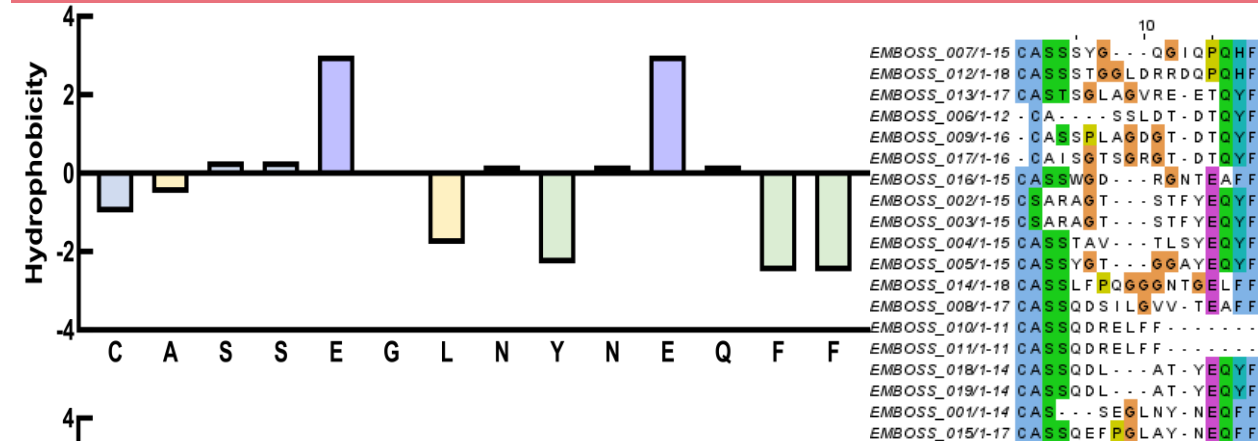
TCR 407



TCR 7939



### CDR3-β Chain



Aliphatic Aromatic Polar Acidic Basic

- AA Residues Colored by ClustalX Conservation

# Identification of Expanded TCR Clonotypes in Peptide

## Variable Sequence Clustering, Motif Generation, and Residue Correlation

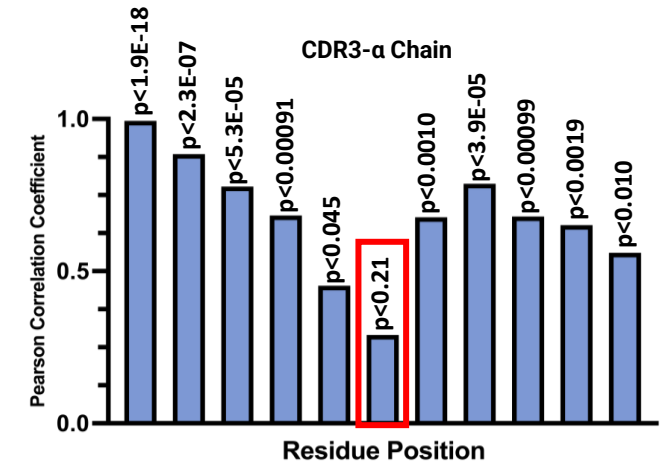
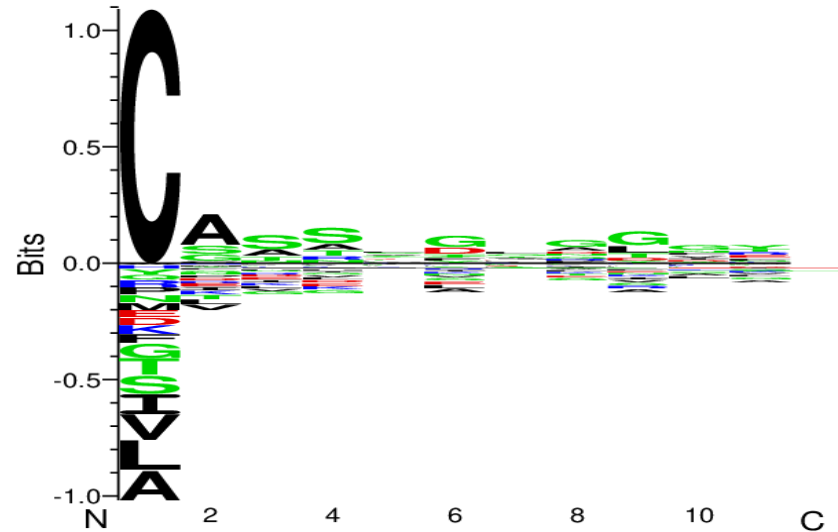
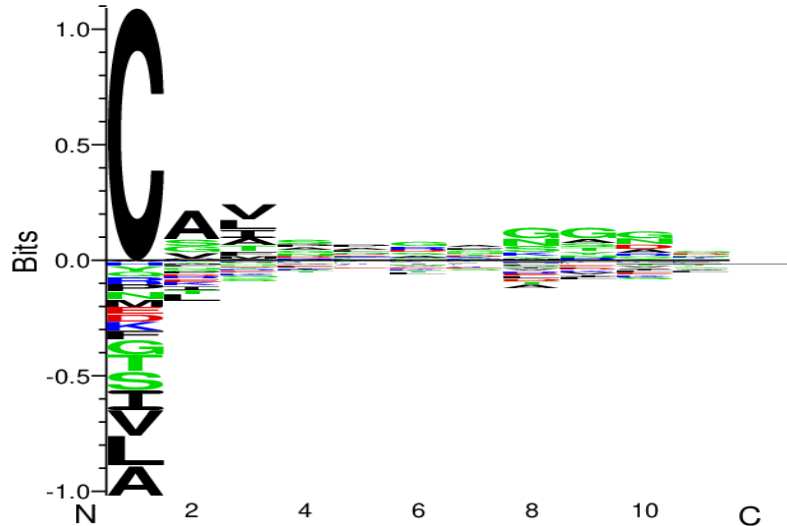
### CDR3- $\alpha$ Chain

### CDR3- $\beta$ Chain

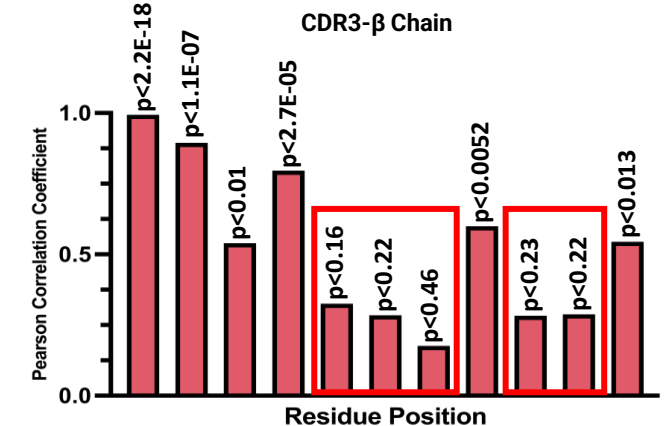
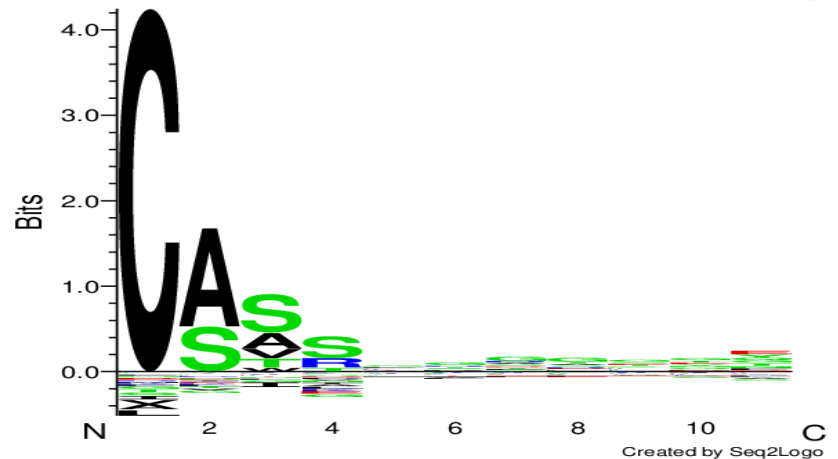
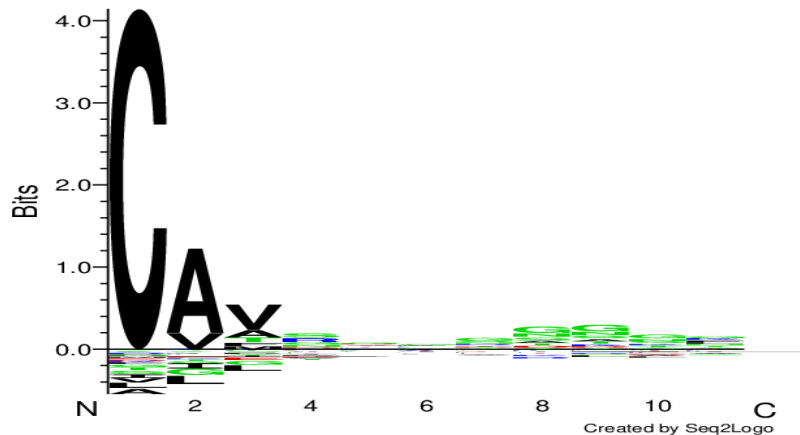
### Position Based Pearson Correlation

Bauer et al., 2022 (*Nature Comm.*)

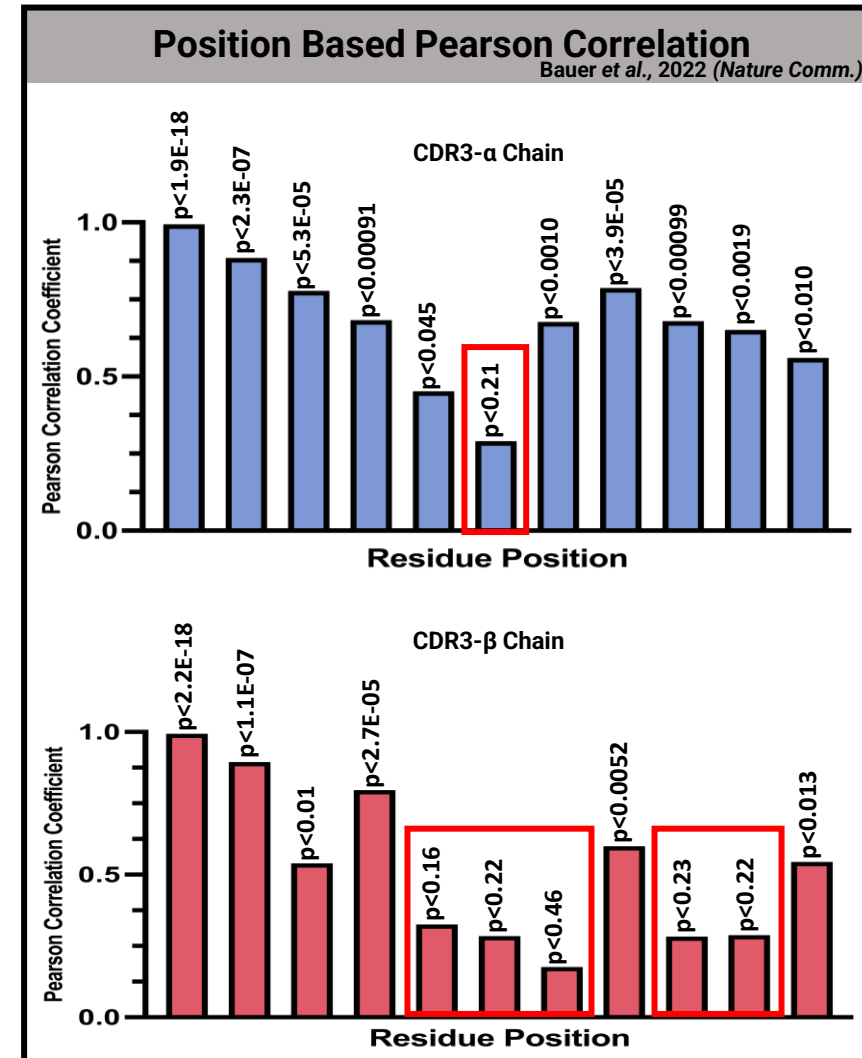
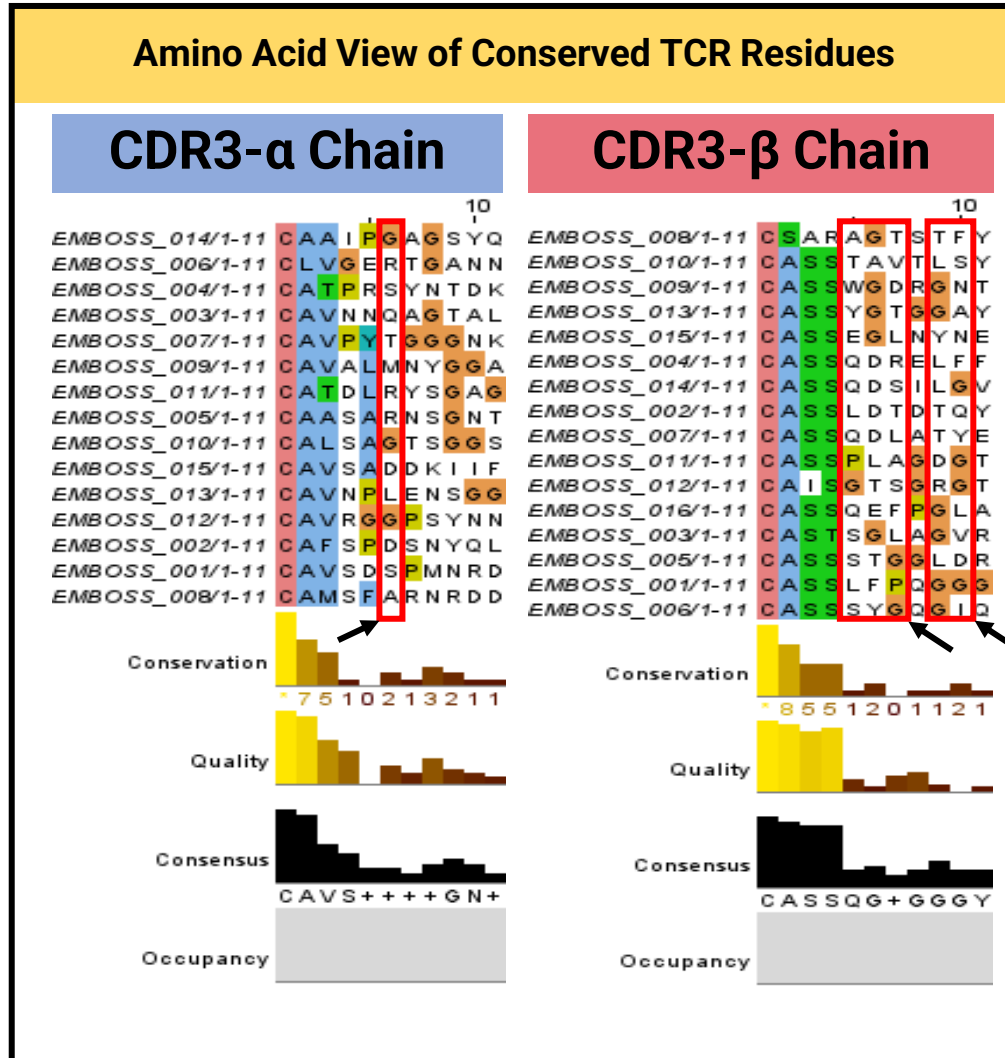
Top Expanded TCRs



Naïve TCRs



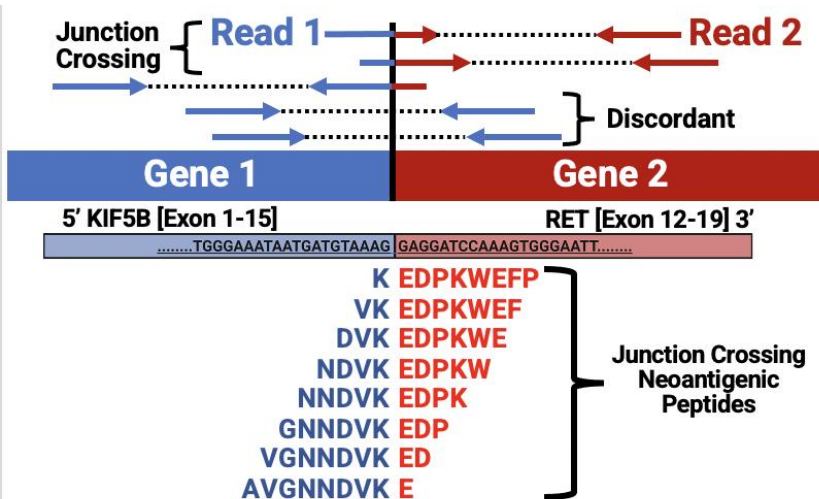
# Characterization of Top Expanded TCR Clonotypes



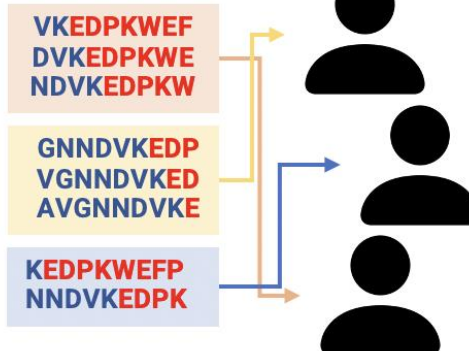
Variable Residue Glycines  
in Expanded T-Cells  
• de Greef, et al., 2021  
(PNAS)  
• Yassai, et al., 2017 (J.  
Immunol.)

- AA Residues Colored by ClustalX Conservation

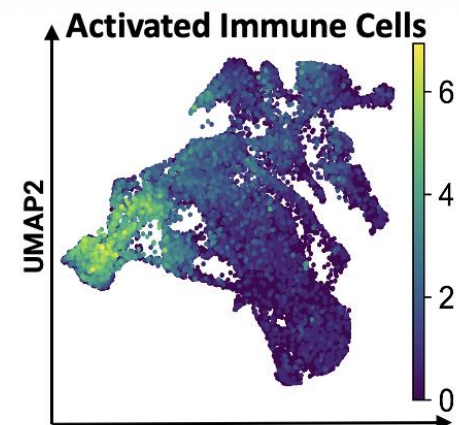




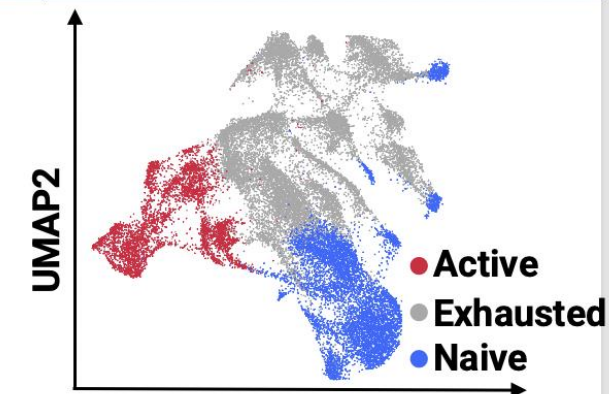
## Personalized Vaccination



## Characterize Immune Response

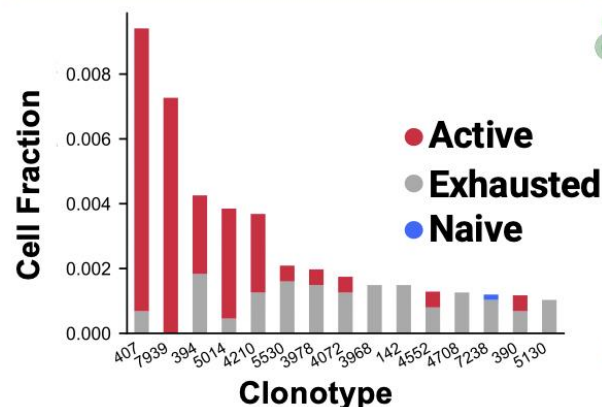


## Define Immune Populations

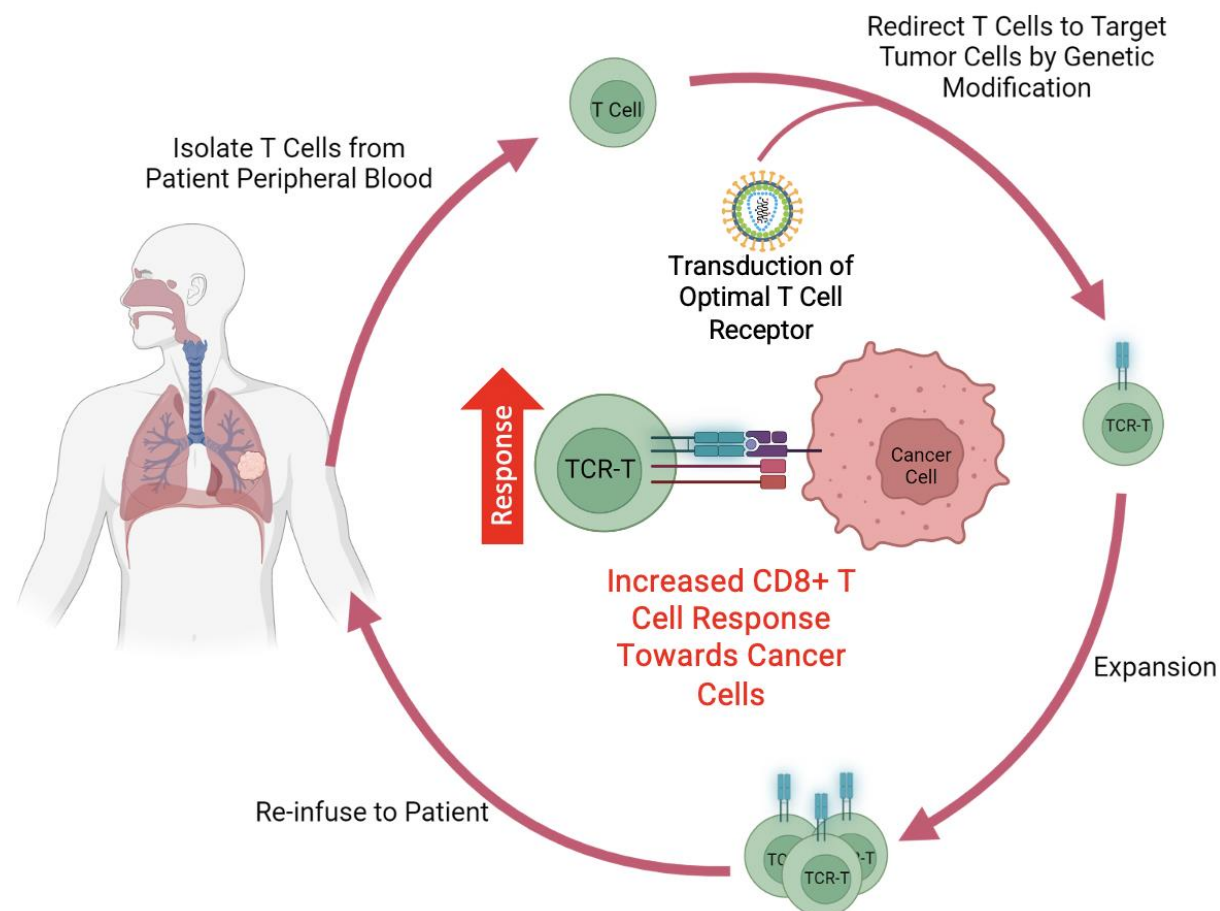
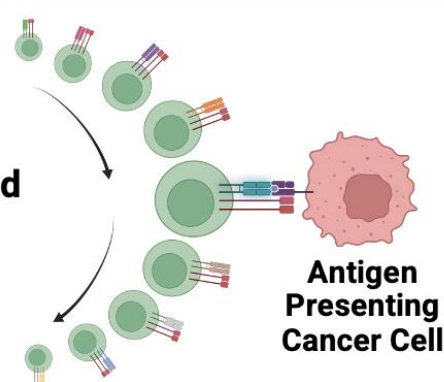


## UMAP1

### Profile T Cell Receptor (TCR) Landscape



## TCR Selection



75 Patients

- 25 HR+
- 25 HER2+
- 25 TNBC

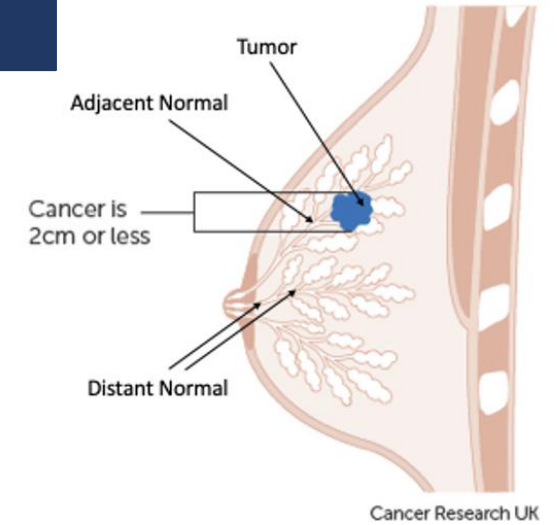
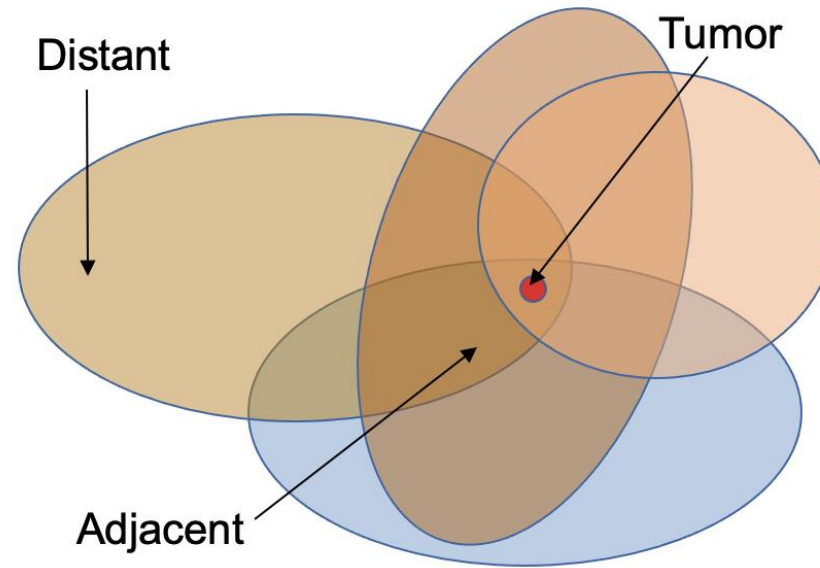
4 Sites

- Tumor
- Adjacent Normal
- 2 Distant Normal

4 Normal Controls

- ~25M read/sample

## Breast Cancer Prevention 4-Site Sequencing



## Phase I Study of a Personalized Breast Cancer Prevention Vaccine

Woman with precancer/at risk for cancer

- Atypia
- Lobular neoplasia

Sequencing of breast tissue

Bioinformatics analysis to identify the gene targets

Manufacture the vaccine against these gene targets

Vaccinate

Test whether vaccine generated immune response

# Fusion Discovery Pipeline

1. 75 Tumor samples from Sporadic 360sample cohort.
2. 43 BRCA1\_2 mutated samples



Filtered for Driver Gene(COSMIC database) associated Gene Fusions only



Removed Fusion Genes detected in 21 Normal Samples



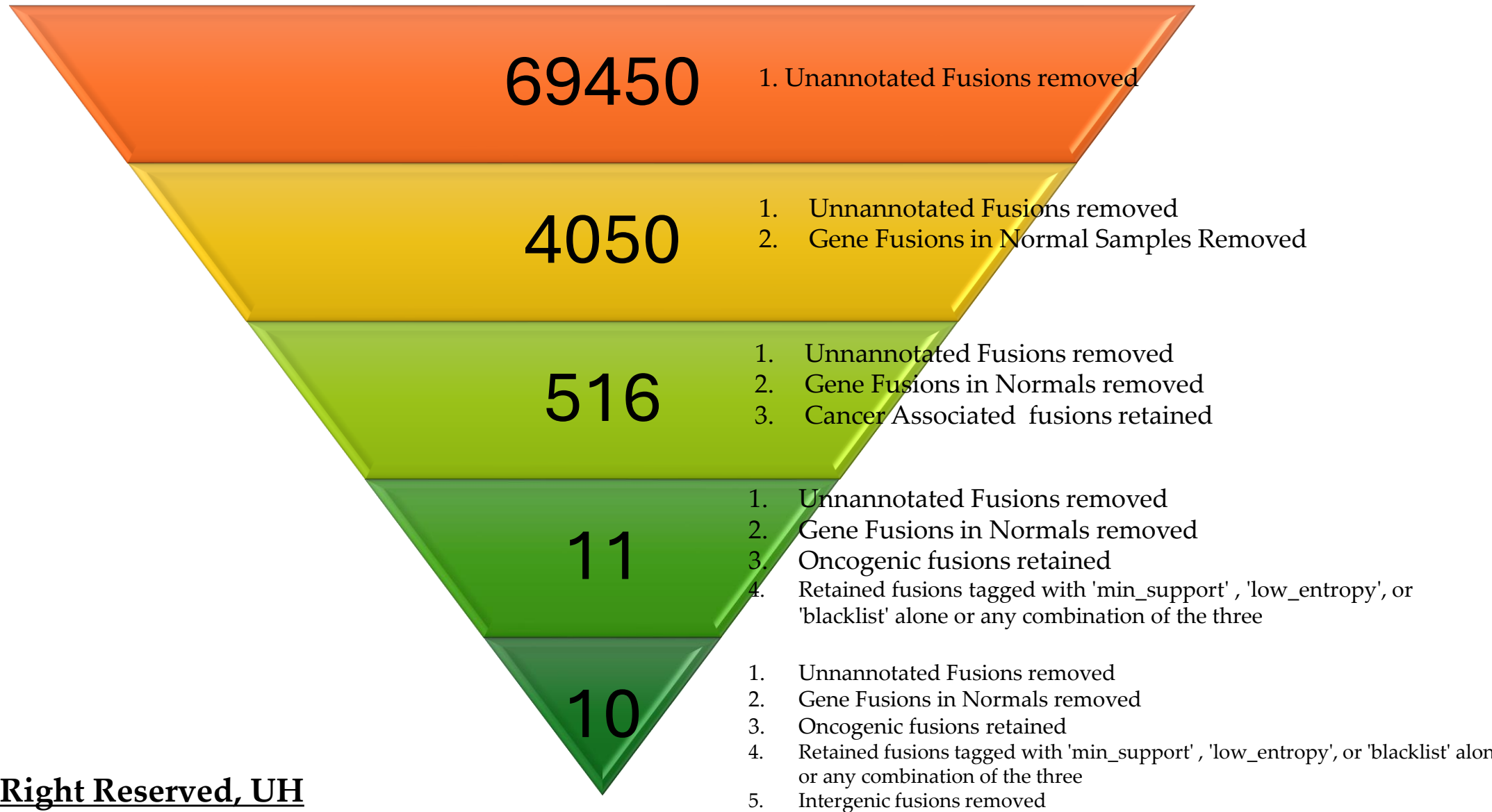
Overlapping Fusion Genes were determined between the two datasets(shown above)



Overlapping Fusion Genes across subtypes were determined between the two datasets(shown above)

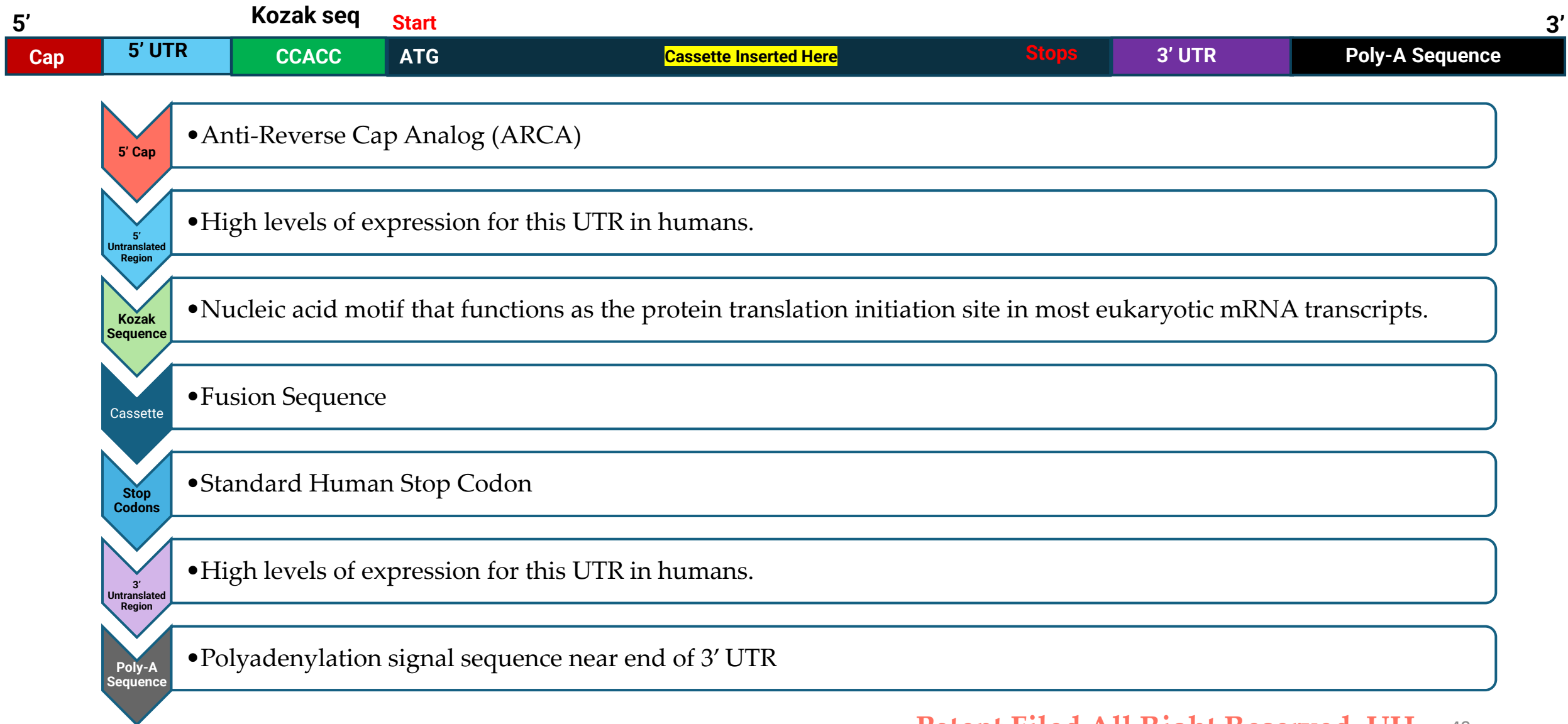


# Overlapping fusions across 75 Tumors and BRCA1\_2 High Risk Dataset





# mRNA Vaccine for Treatment & Prevention of Cancer



# Peptide and mRNA Vaccine Development Team

**Sakuni  
Rankothgedera**



Lead Scientist  
Single Cell and Spatial  
Transcriptomics  
Platform

**CD74- NRG1**

Lung Cancer  
Ovarian Cancer

**Aaranyah  
Kandasamy**

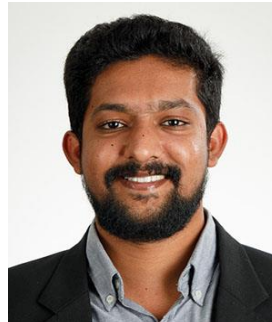


Lead Scientist  
PDX -Actionable Mutation  
Discovery Platform

**ESR1-PRKN**

Breast Cancer

**Shiyanth  
Thevasagayampillai**



Lead Scientist  
mRNA Vaccine  
Discovery Platform

**FGFR3-TACC3**

Brain Cancer  
Lung Cancer  
Breast Cancer  
Pancreatic Cancer  
Ovarian Cancer

**Dilshan Adikari**



NGS Specialist  
RNA & DNA  
Sequencing Platform

**FGFR3-KHSRP**

Pancreatic Cancer  
Brain Cancer  
Ovarian Cancer  
Urothelial Cancer

**Cole Woody**

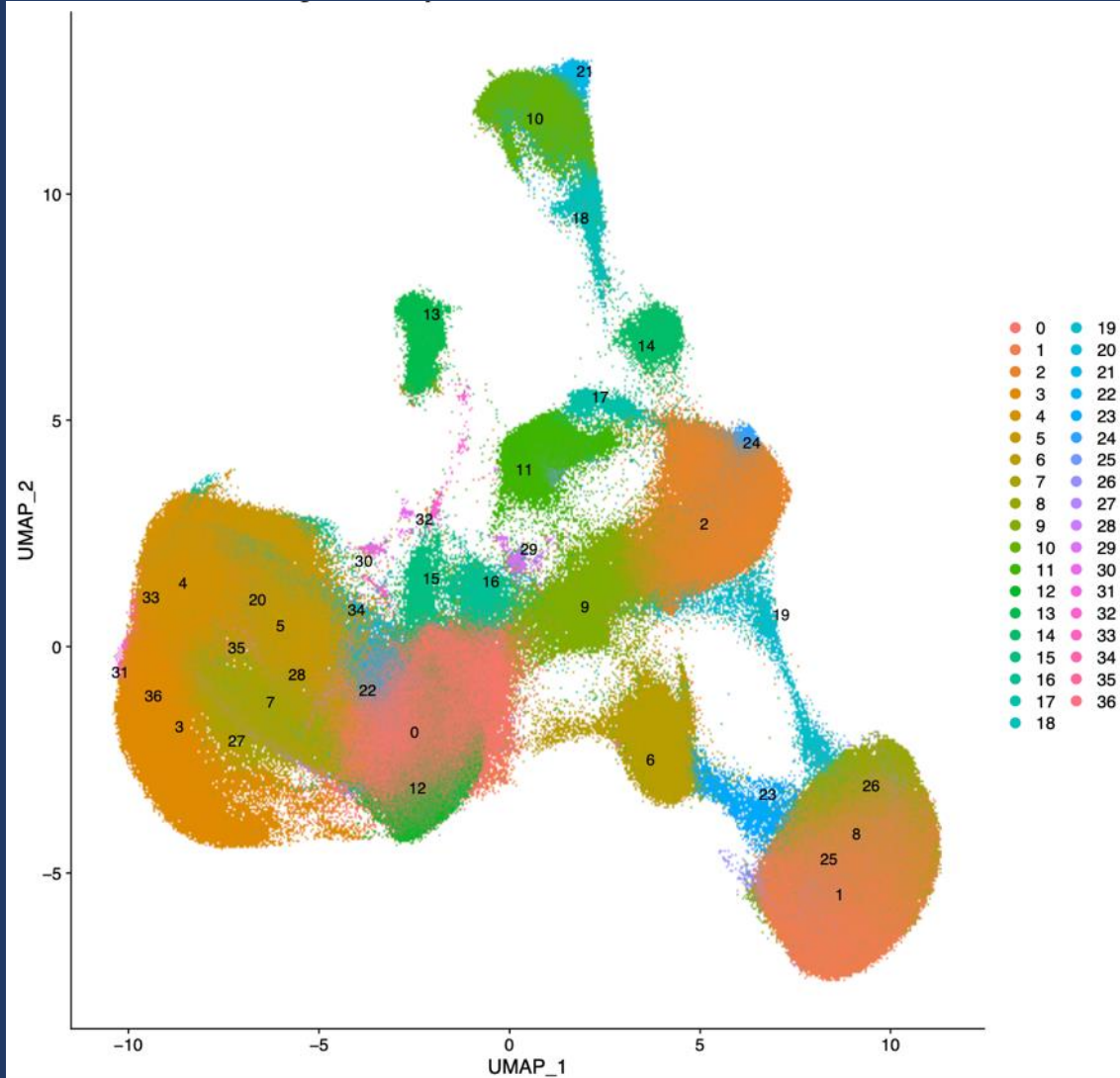


*In silico* fusion  
validation platform

**HNRNPM-CCNE1**

Ovarian Cancer

# Sequencing a Million Cells from over a Hundred Brains from Opioid Use Disorder (OUD) Patients



1 Million Cells Profiled From Cryopreserved Brain

Hypothalamus

~400,000 Cells

Thalamus

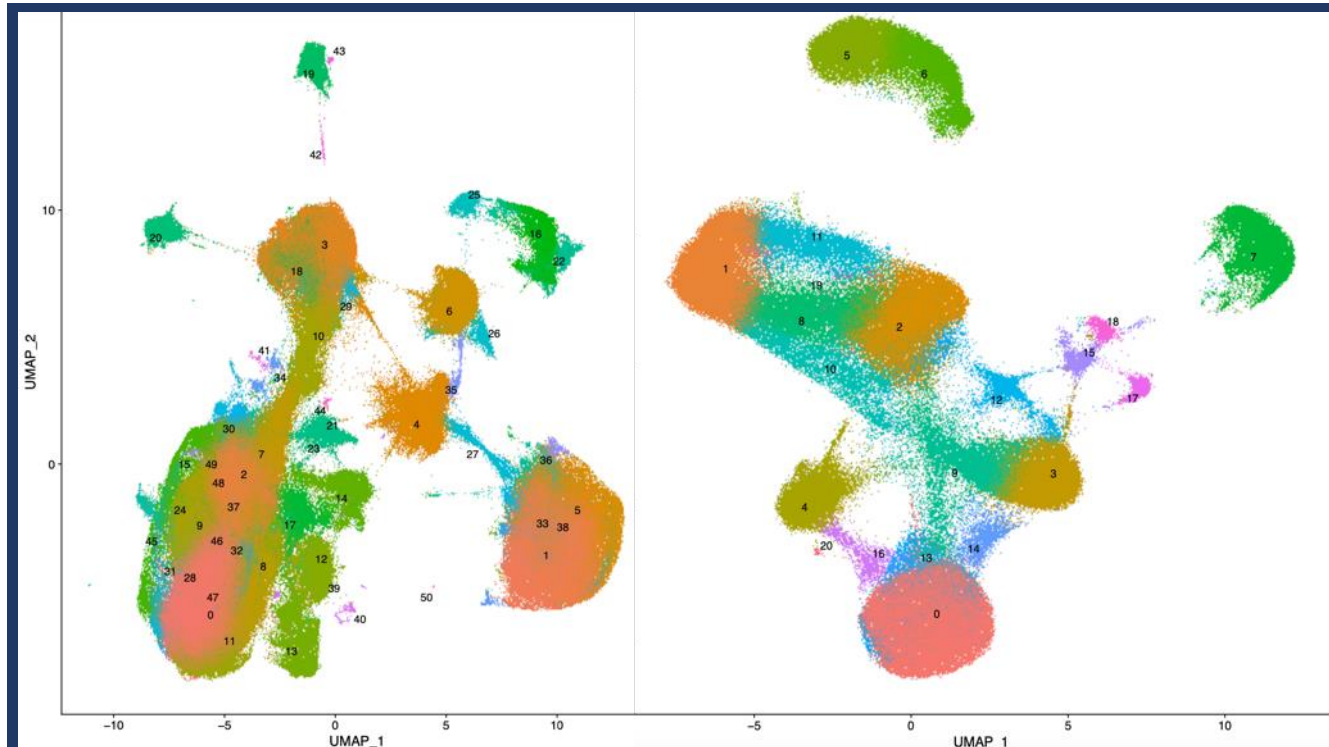
~300,000 Cells

Globus Pallidus

~300,000 Cells

Hypothalamus

Thalamus



# microRNA Biomarkers for Opioid Addiction Risk

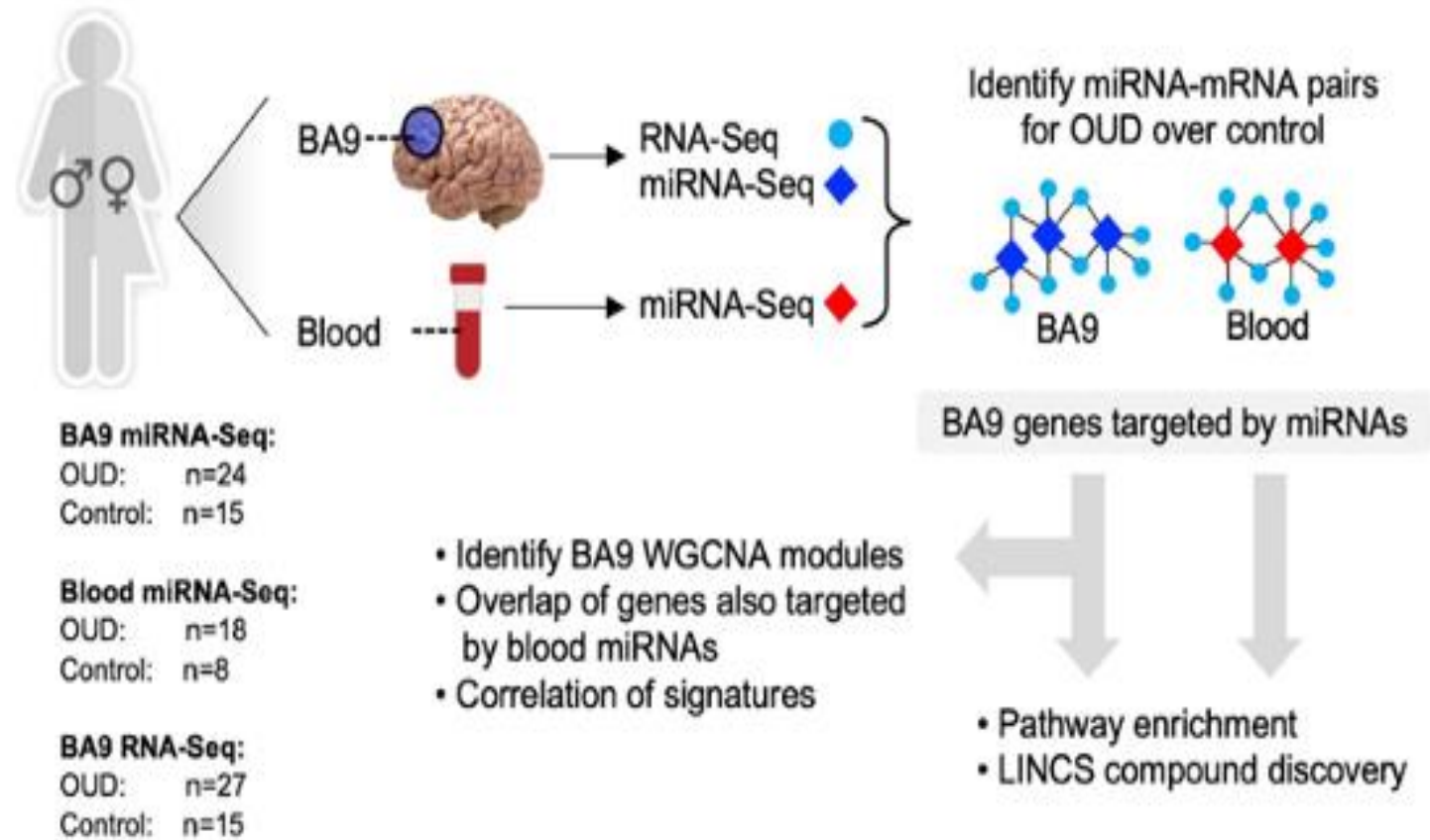


**Thor Loveland**  
Founder and CEO, miRMIND

## miRMIND

### miR-Tracker

- Blood Biomarker
- Predict & prevent death from opioid overdose





# UH Sequencing and Gene Editing Core



## Administration

- Dr. Preethi Gunaratne – Director
- Sakuni Rankothgedera – Lead Scientist
- Aaranyah Kandasamy – Project Manager
- Shiyanth Thevasagayampillai – Manager Analytics Platforms

## Business Office Liaisons

- Thien-Tam Do
- Malinthika Fernando
- Fred McGhee
- Judy Mata
- Rachel Marks



## Core Collaborators

- **MD Anderson Cancer Center**
  1. Dr. Hussein Abbas
  2. Dr. Moran Amit
- **UT Health Science Center**
  1. Dr. Consuelo Walss-Bass – UT Health
- **Baylor College of Medicine**
  1. Dr. Andrew Dinardo
  2. Dr. Cristian Coarfa
- **University of Houston**
  1. Dr. Beau Alward – UH
  2. Dr. Mingfu Wu – UH
  3. Dr. Ashok Kumar – UH
  4. Dr. Sang-Hyuk Chung – UH
  5. Dr. Tasneem Bawa – UH
  6. Dr. Robert Schwartz – UH
  7. Dr. Meghana Trivedi – UH
- **Columbia University, NY**
  1. Dr. Emily Mace
- **Augusta University, GA**
  1. Dr. Huabo Su – Augusta University
  2. Dr. Hong Shi – Augusta University
  3. Dr. Xiaochun Long – Augusta University
- **University of South Florida, FL**
  1. Dr. Wanling Xuan – USF
- **Texas Tech University**
  1. Dr. Naima Moustaid-Moussa – TTU
- **Texas Childrens Hospital**
  1. Dr. Gabriel Loor
  2. Dr. Lourdes Chacon